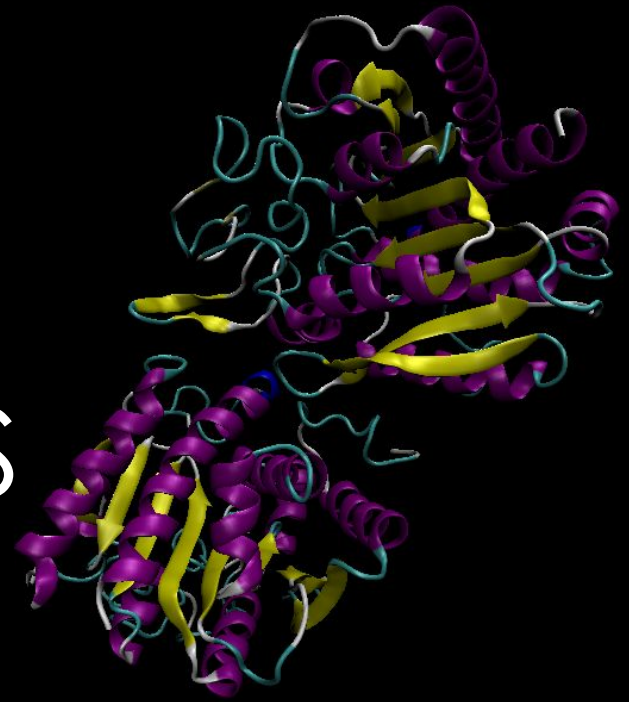


CARBOXYPEPTIDASES



Esther Colomé
Kristin Fichna
Carla Mateo
Maria Padial

INDEX

1. Introduction
 - a. Peptidases
 - b. Classification
 - i. PFAM
 - ii. MEROPS
 - iii. SCOP
2. Carboxypeptidases evolution
3. Structural analysis
 - a. Rossmann fold
 - b. Catalytic site
 - c. Pro-carboxypeptidases
4. Carboxypeptidase inhibitors
5. Conclusion
6. References

PEPTIDASES

- Proteolytic enzymes
- Broad spectrum of functions
- Medicine
- Biological processes

**Dual classification
depending on the
cleavage they perform:**

Endopeptidases

Exopeptidases →

Carboxypeptidases

**Residue that is essential
for catalysis:**

- Serine proteases
- Cysteine proteases
- Aspartic acid proteases
- Threonine proteases
- Glutamic acid proteases
- Metalloproteases

CARBOXYPEPTIDASES CLASSIFICATION

By active site mechanism

Metalloprotease



Enzymes that use a metal in the active site

Serine protease



Enzymes that use a serine in the active site

Cysteine protease



Enzymes that use a cysteine in the active site

By substrate preference

Carboxypeptidase A



Aromatic/aliphatic residues

Carboxypeptidase B



Basic residues

MEROPS CLASSIFICATION

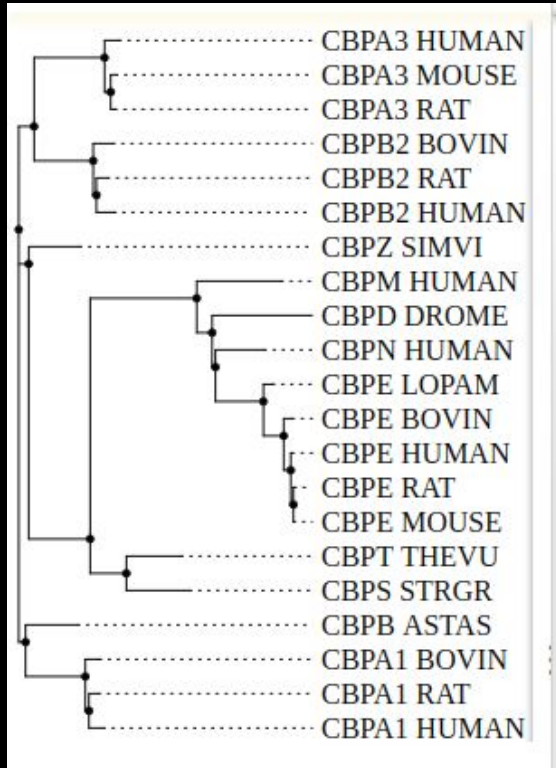
Clan	Family	Subfamily	Subfamily type peptidase
MC	M14	M14A	Carboxypeptidase A
		M14B	Carboxypeptidase E
		M14C	Gamma-D-glutamyl-(L)-meso-diaminopimelate peptidase I
		M14D	Cytosolic carboxypeptidase 6

<https://www.ebi.ac.uk/merops/cgi-bin/famsum?family=M14>

SCOP CLASSIFICATION

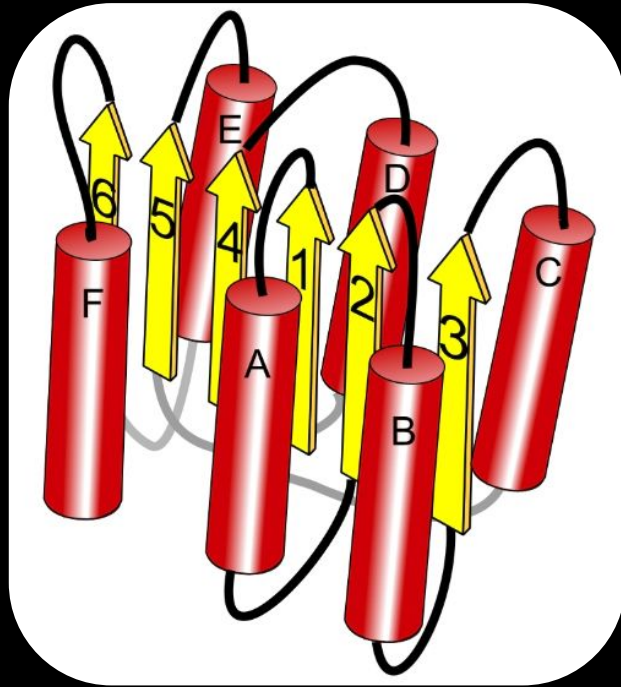
Class	Fold	Superfamily	Family
Alpha and beta proteins (a/b)	Phosphorylase /hydrolase-like	Zn-dependent exopeptidases	Pancreatic carboxypeptidases
			Carboxypeptidase T
			Leucine aminopeptidase, C-terminal domain
			Bacterial dinuclear zin exopeptidases
			FolH catalytic domain-like
			N-acetylmuramoyl-L-alanine amidase-like
			AstE/AspA-like
			Glutaminyl-peptide cyclotransferase-like
			FGase-like

CARBOXYPEPTIDASES EVOLUTION

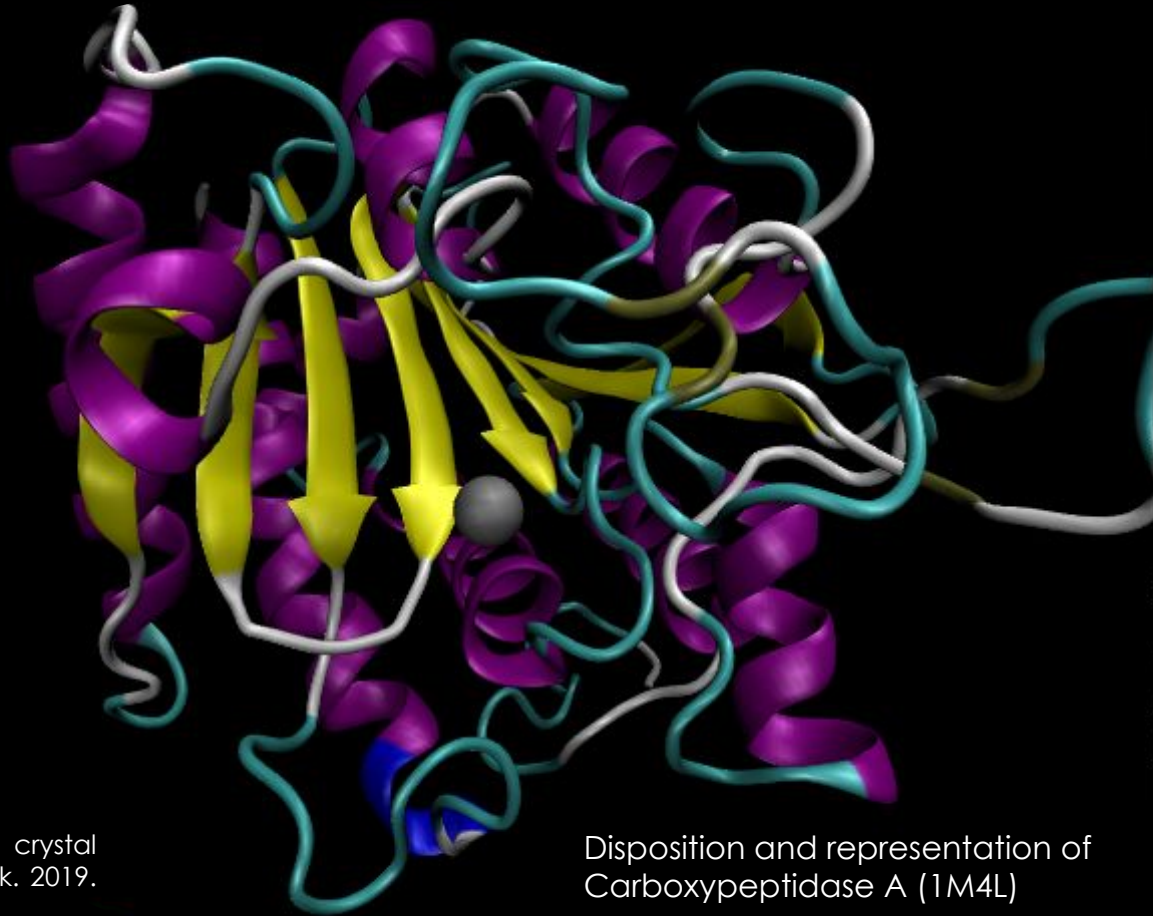


- Phylogenetic tree
- Carboxypeptidases A and B
- Mast cells
- Regulatory Carboxypeptidases

Scaffold

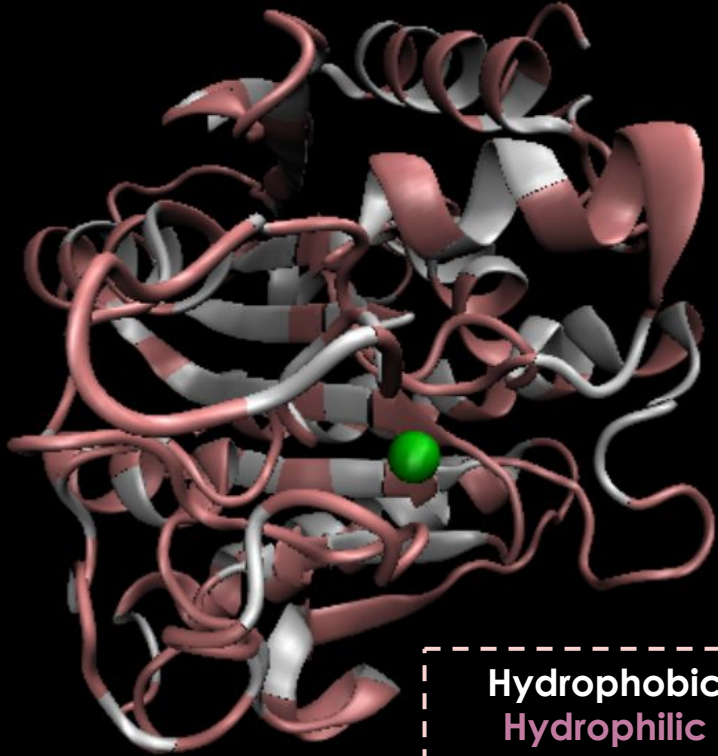


Europe P. Phaser - a "stunning" method for solving crystal structures ‹ Quips ‹ PDBe ‹ EMBL-EBI [Internet]. Ebi.ac.uk. 2019. Available from: <http://www.ebi.ac.uk/pdbe/quips?story=Phaser>



Disposition and representation of Carboxypeptidase A (1M4L)

Bovine Carboxypeptidase A



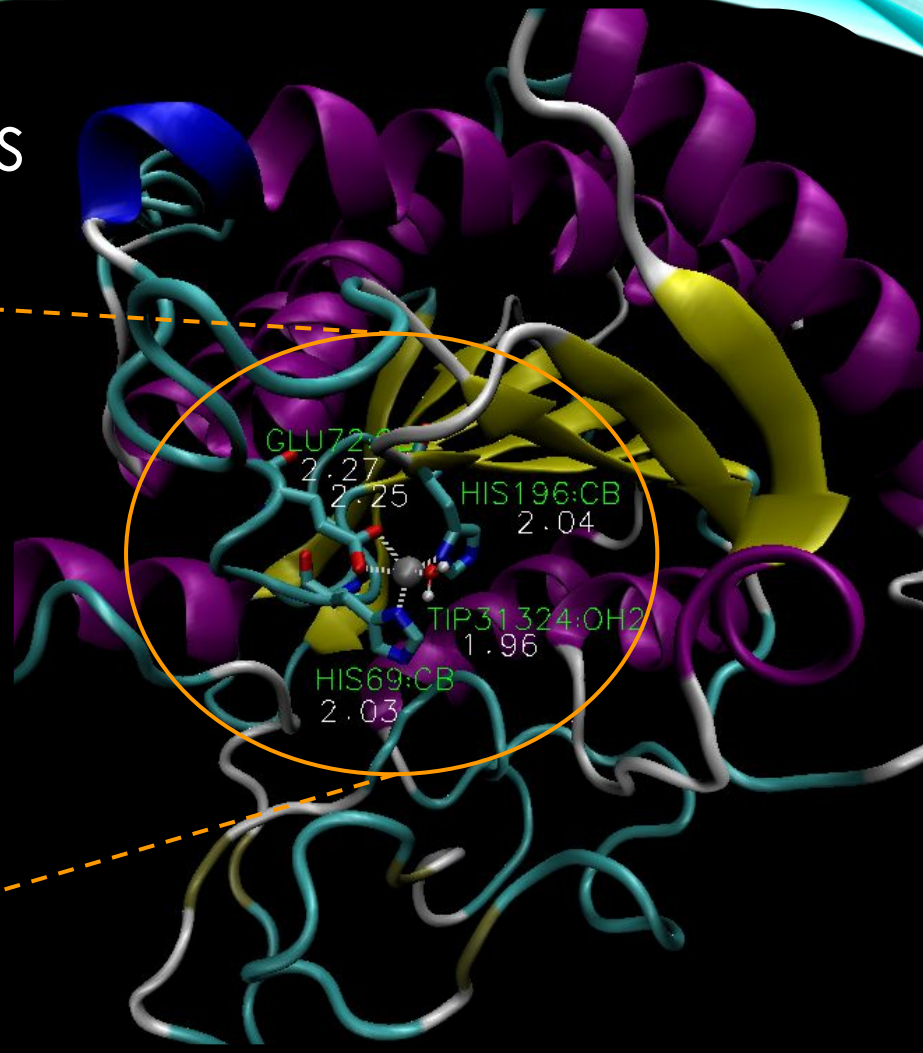
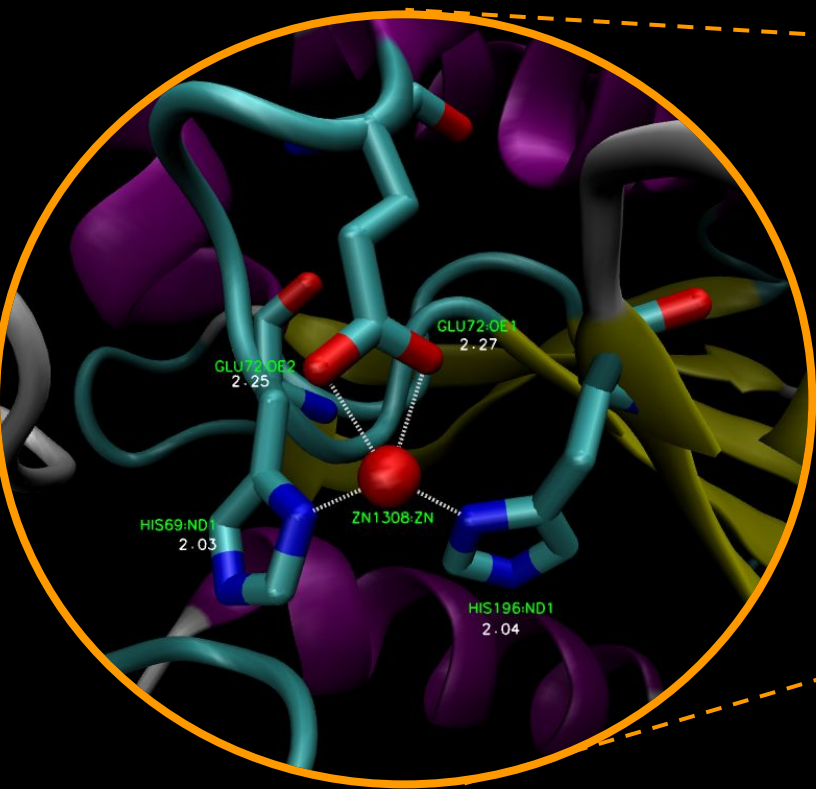
Hydrophobic residues
Hydrophilic residues
Zinc ion





CATALYTIC SITE

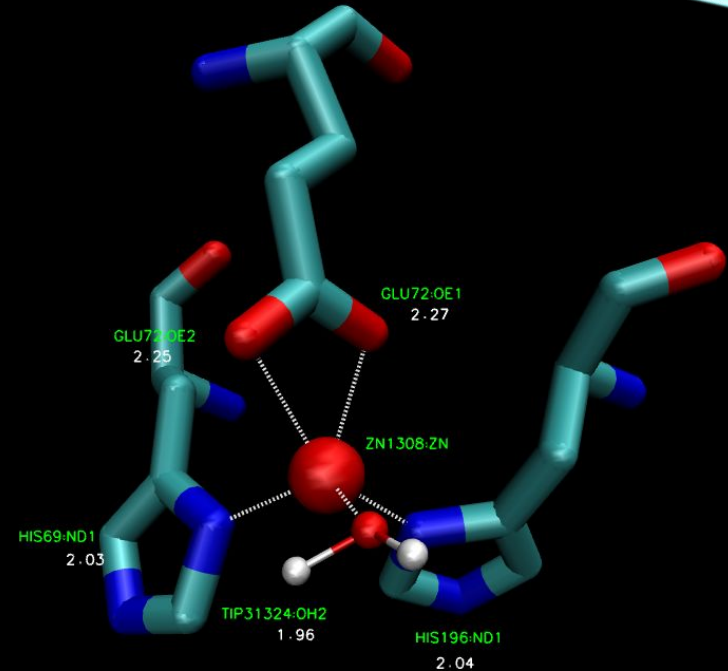
Zn coordination residues



Zn coordination

His 60 Glu 72

CBPA2_HUMAN	HAREWVTQATALWTANKIVSDYGKDPS
CBPA1_RAT	HSREWVTQASGVWFAKKITKDYGQDPT
CBPA1_MOUSE	HSREWVTQASGVWFAKKITKDYGQEPT
CBPA1_BOVIN	HSREWITQATGVWFAKKFTEDYGQDPS
CBPA1_HUMAN	HSREWVTQASGVWFAKKITQDYGQDAA
CBPA1_PIG	HSREWVTQASGVWFAKKITEDYGQDPA
	::*:*:*:*:*:*:*:*:*:*:*:*:*:

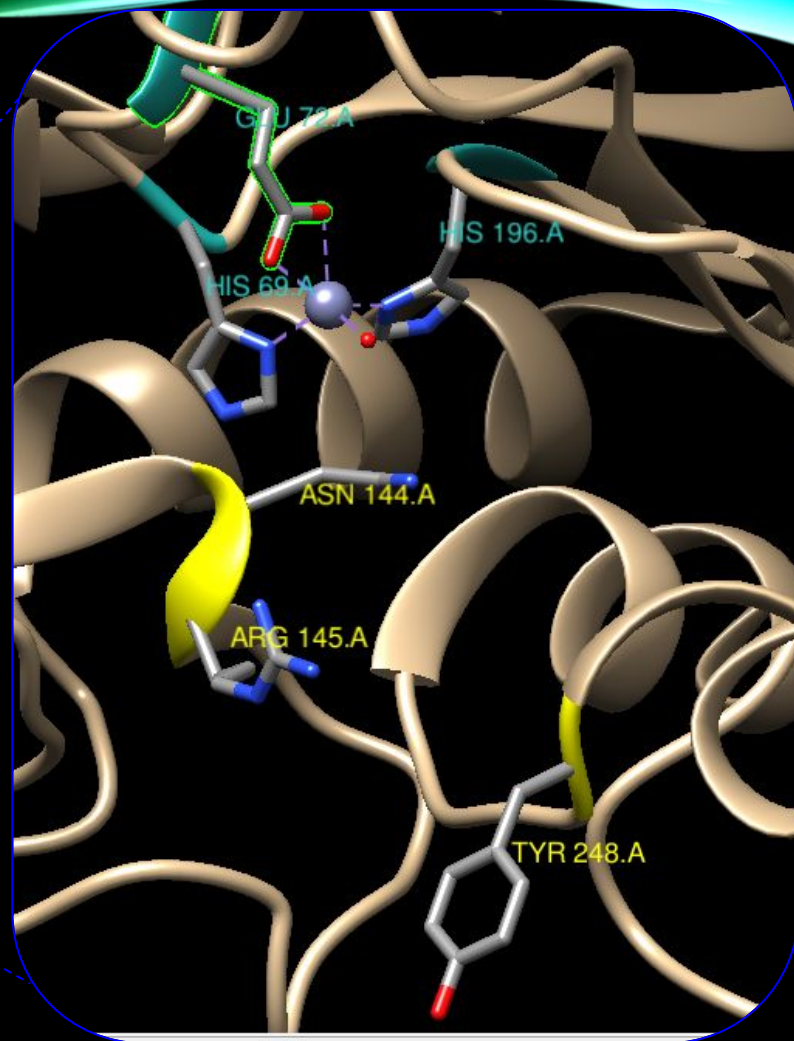
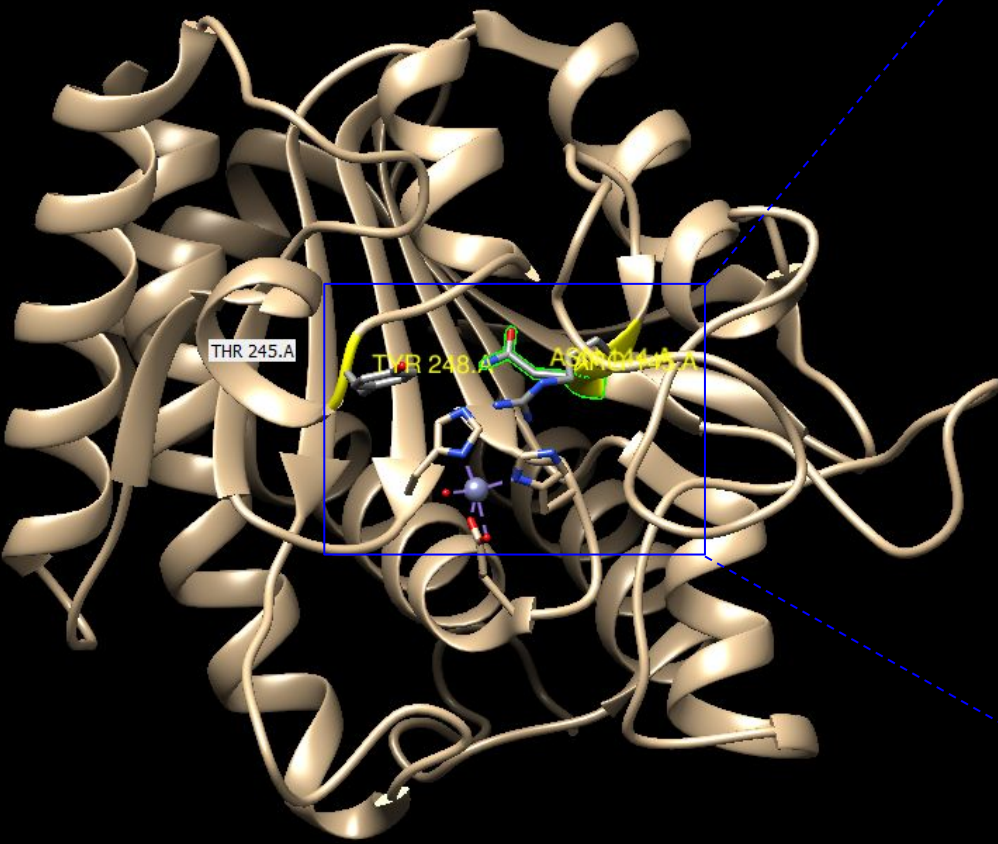


His 196

CBPA1_RAT	TRSHTQGS LCVGVDPNRNWDAGFGMAGASSNPCSE TYRGKFPNSEVEVKSI VDFVTS HGN
CBPA1_MOUSE	TRSHTEGSLCVGVDPNRNWDAAF GMPGASSNPCSE TYRGKFPNSEVEVKSI VDFVTS HGN
CBPA1_HUMAN	TRSHTAGSLCIGVDPNRNWDAGFGLSGASSNPCSE TYHGKFANSEVEVKSI VDFVKD HGN
CBPA1_PIG	TRSRTSGSFCVGVDPNRNWDAGFGGAGASSNPCSE TYHGKFPNSEVEVKSI VDFVND HGN
CBPA1_BOVIN	TRSVTSSSLCVGV DANRNWDAGFGKAGASSSPCSE TYHGKYANSEVEVKSI VDFVKD HGN
CBPA2_HUMAN	TRSKVSGSLCVGVDPNRNWDAGFGGPGASSNPCSDSYHGPSANSEVEVKSI VDFIKSHGK
	*** * . * . * * * * * * * * * * * * * * * * . * * * * * * * * * * * * * * * . * *

Residues His 69, His 196 and Glu 72 are conserved among species.

S1' subsite



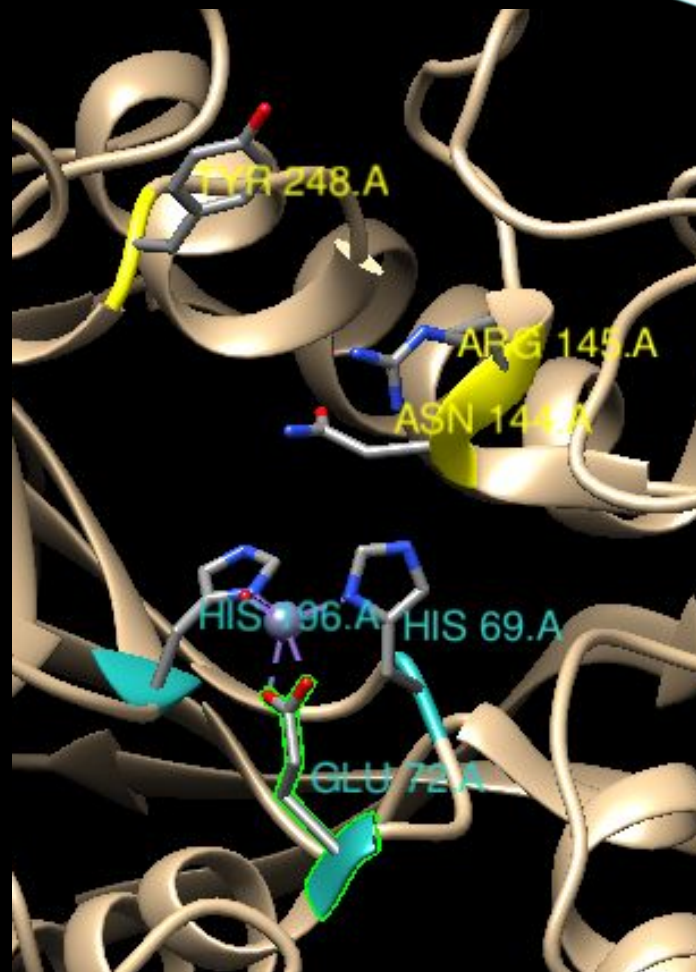
S1' subsite

Asn 144-Arg 145

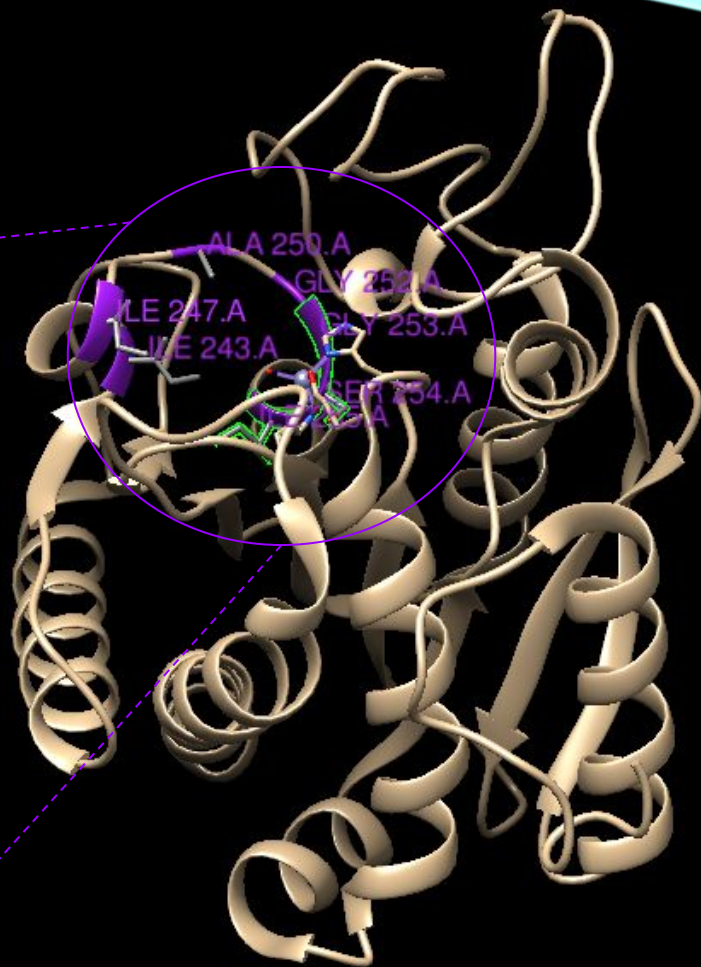
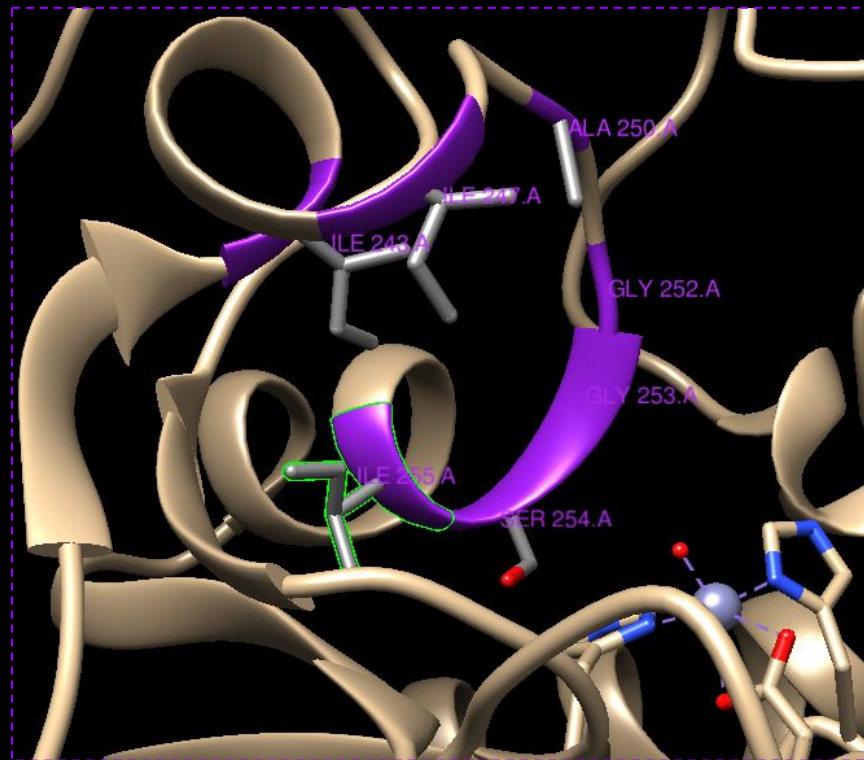
CBPA1_RAT	TRSHTQGSLCVGVDPNRNWDAGFGMAGASSNP
CBPA1_MOUSE	TRSHTEGSLCVGVDPNRNWDAAFGLMPGASSNP
CBPA1_HUMAN	TRSHTAGSLCIGVDPNRNWDAGFGLSGASSNP
CBPA1_PIG	TRSRTSGSFCVGVDPNRNWDAGFGGAGASSNP
CBPA1_BOVIN	TRSVTSSSLCVGVDPNRNWDAGFGKAGASSNP
CBPA2_HUMAN	TRSKVSGSLCVGVDPNRNWDAGFGGPGASSNP
	*** . . *:***.*****.*** .*****.*

LLMFPYGYKCTKLDDFDELSEVAQKAAQSLRSLHGTTYKVGPICSVIY
LLLYPYGYTSEPAPDQAELDQLAKSAVTALTSLHGTYKFGYGSIIIDTIY
LLLYPYGYTSEPAPDKEELDQLAKSAVTALTSLHGTYKFGYGSIIIDTIY
LLLYPYGYTTQSIPDKTELNQVAKSAVEALKSLYGTSTYKYGSIIITTIY
LLMYPYGYKTEPVPDQDELQLSKAAVTALASLYGTGFNYGSIIKAIY
LLLYPYGYKTEAPADKDELQISKSAAVATSLYGTGFQYGSIIITTIY
::***. * **:::*. :* **::**::: * * .**

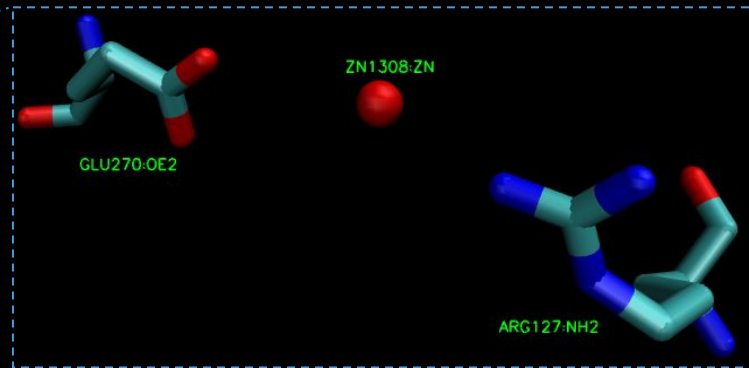
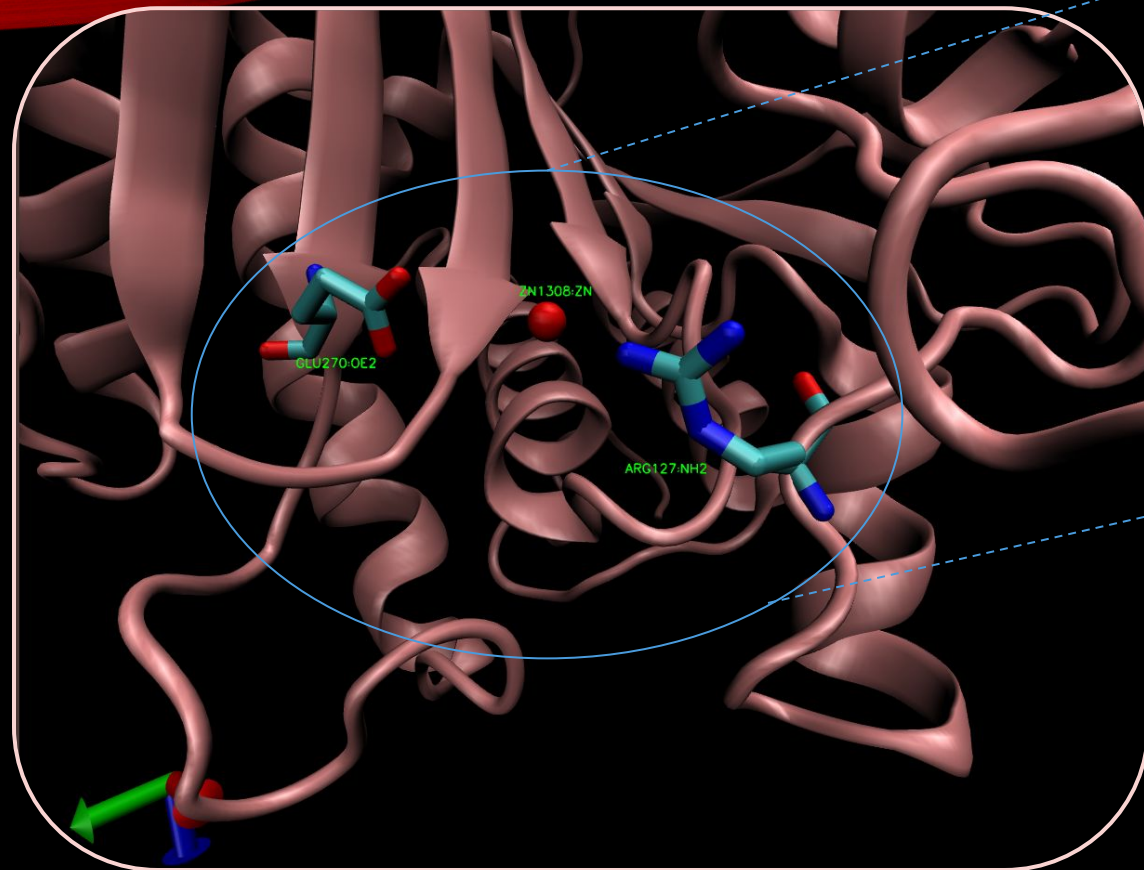
Tyr 248



Hydrophobic Pocket



S1 subsite



S1 Subsite

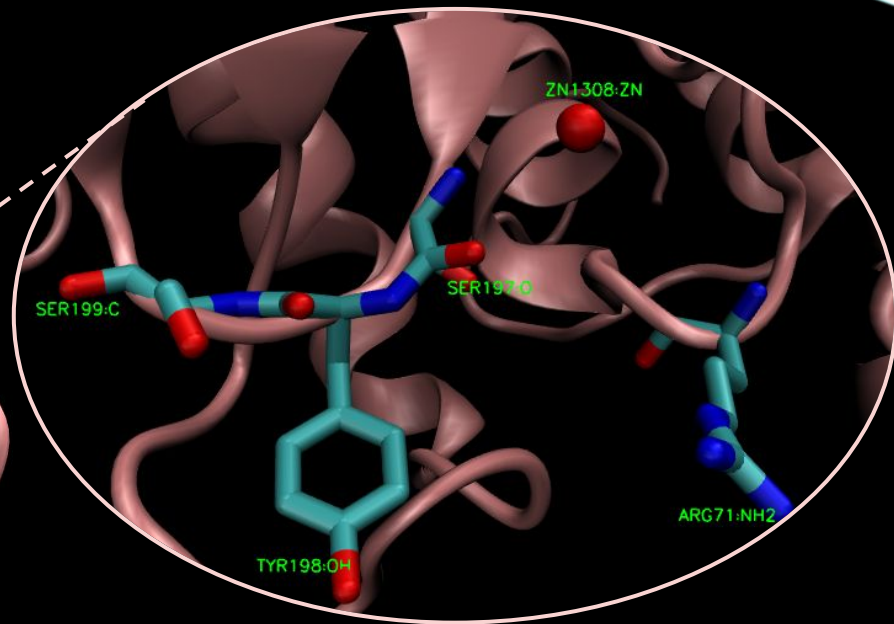
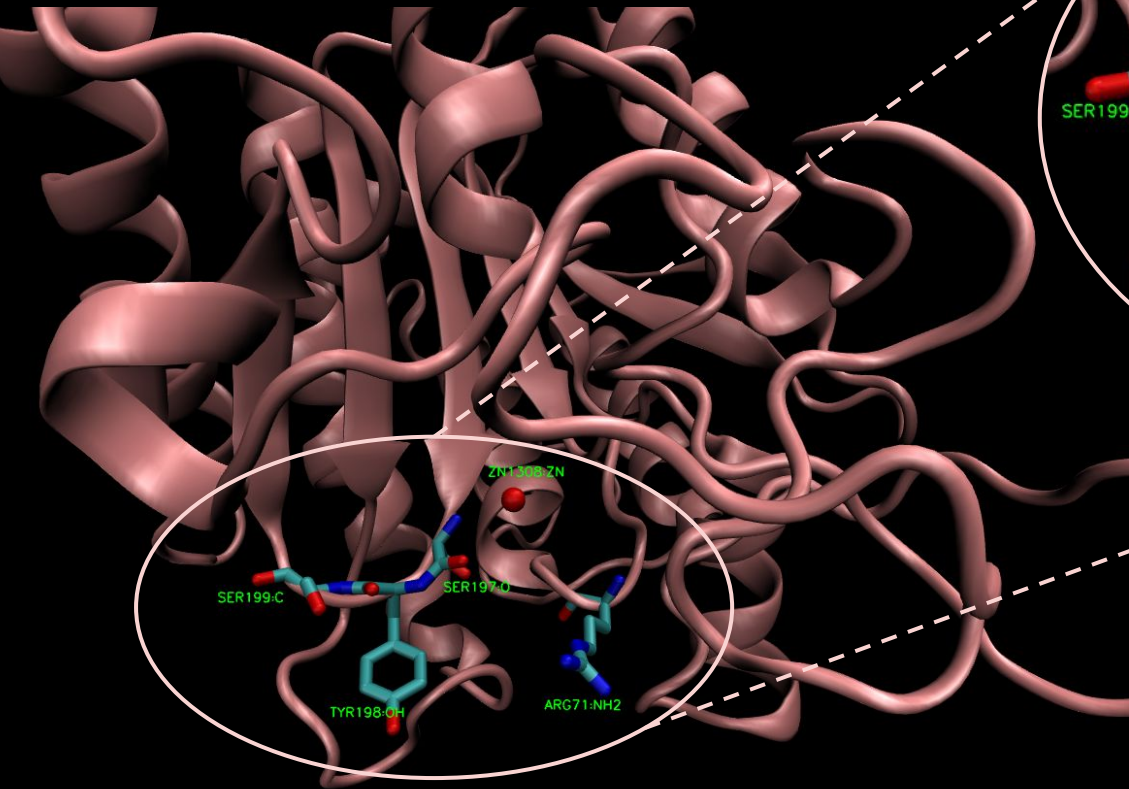
Arg 127

Species	Sequence
CBPA2_HUMAN	HAREWVTQATALWTANKIVSDYGKDPSITSILDALDIFLLPVTNPDGYVFSQTKNRMWRK
CBPA1_RAT	HSREWVTQASGVWFAKKITKDYGQDPTFTAVLDNMDIFLEIVTNPDGFAYTHKTNRMWRK
CBPA1_MOUSE	HSREWVTQASGVWFAKKITKDYGQEPTLTAILDNMDIFLEIVTNPDGFVYTHKTNRMWRK
CBPA1_BOVIN	HSREWITQATGVWFAKKFTEDYGQDPSFTAILDSMDIFLEIVTNPDGFAFTHSQNRLWRK
CBPA1_HUMAN	HSREWVTQASGVWFAKKITQDYGQDAAFTAILDTLDIFLEIVTNPDGFAFTHSTNRMWRK
CBPA1_PIG	HSREWVTQASGVWFAKKITEDYGQDPAFTAILDNLDIFLEIVTNPDGFAFTHSENRMWRK
	*:***:***:..* *:*.:.***: :*:*** :***** :*****:..:.. *:**

Glu 270

CBPA2_HUMAN	QASGGSIDWSYDYGIKYSFAFE	LRDTGRYGFLLPARQILPTAEETWLGLKAIMEHVRDHP
CBPA1_RAT	QASGSTIDWTYSQGIKYSFTFE	LRDTGLRGFLLPASQIIPTAEETWLALLTIMDHTVKHP
CBPA1_MOUSE	QASGSTIDWTYSQGIKYSFTFE	LRDTGLRGFLLPASQIIPTAEETWLALLTIMDHTVKHP
CBPA1_BOVIN	QASGGSIDWSYNQGIKYSFTFE	LRDTGRYGFLLPASQIIPTAQETWLGVLTIMEHTLNNL
CBPA1_HUMAN	QASGSTIDWTYSQGIKYSFTFE	LRDTGRYGFLLPASQIIPTAKETWLALLTIMEHTLNHP
CBPA1_PIG	QASGGTIDWTYNQGIKYSFSFE	LRDTGRYGFLLPASQIIPTAQETWLALLTIMEHTLNHP
	****.:.****.*.*****:*****	***** **;****:*****.: **:*.:

S2 Subsite



S2 Subsite

Arg 71

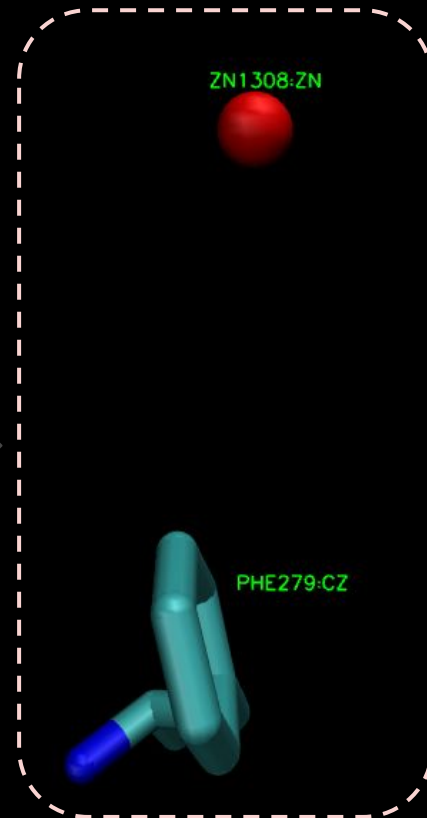
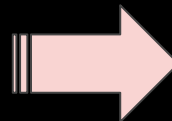
CBPA2_HUMAN HAREWVTQATALWTANKIVSDYGKDPSITSILDALDIFLLPVTNPDGYVFSQTKNRMWRK
CBPA1_RAT HSREWVTQASGVWFAKKITKDYGQDPTFTAVLDNMDFLEIVTNPDGFAYTHKTNRMWKR
CBPA1_MOUSE HSREWVTQASGVWFAKKITKDYGQEPTLTAILDNMDIFLEIVTNPDGFVYTHKTNRMWKR
CBPA1_BOVIN HSREWITQATGVWFAKKFTEDYGGQDPSFTAILDSMDIFLEIVTNPDGFAFTHSQNRLWRK
CBPA1_HUMAN HSREWVTQASGVWFAKKITQDYGDAAFTAILTDLTDLIFLEIVTNPDGFAFTHSTNRMWKR
CBPA1_PIG HSREWVTQASGVWFAKKITEDYGGQDPAFTAILDNLDFLEIVTNPDGFAFTHSENRMWRK

*:

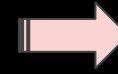
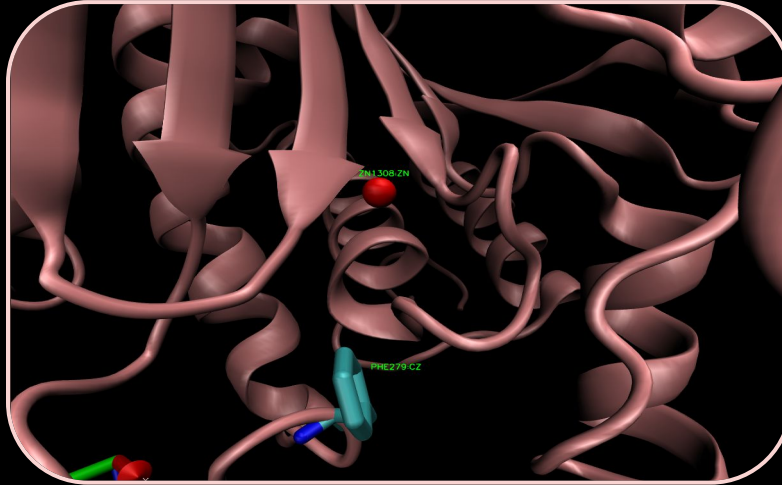
Ser 197/Tyr 198/Ser 199

CBPA2_HUMAN	VKAFITLH	SYSQLLMFPYGYKCTKLDDFDELSEVAQKAAQSLRSLHGTKYKVGPICSVIY
CBPA1_RAT	IKAFISIH	SYSQLLLYPYGYTSEPAPDQAEQLAKSAVTALTSLHGTKFKYGSIIDTIY
CBPA1_MOUSE	IKAFISIH	SYSQLLLYPYGYTSEPAPDKEELDQLAKSAVTALTSLHGTKFKYGSIIDTIY
CBPA1_BOVIN	FKAFLSIH	SYSQLLLYPYGYTTQSIPDKTELNQVAKSAVEALKSLYGTSYKYGSIIITTIY
CBPA1_HUMAN	IKAFISIH	SYSQLLMYPYGYKTEPVPDQDELDQLSKAAVTALASLYGTFKNYGSIIKAIY
CBPA1_PIG	IKAFISIH	SYSQLLLYPYGYKTEAPADKDELDQISKSAVAALTSLYGTFQYGSIIITTIY
	.***:::*	*****:::****. * **:::~*.* **:::~*.* **

S3 subsite

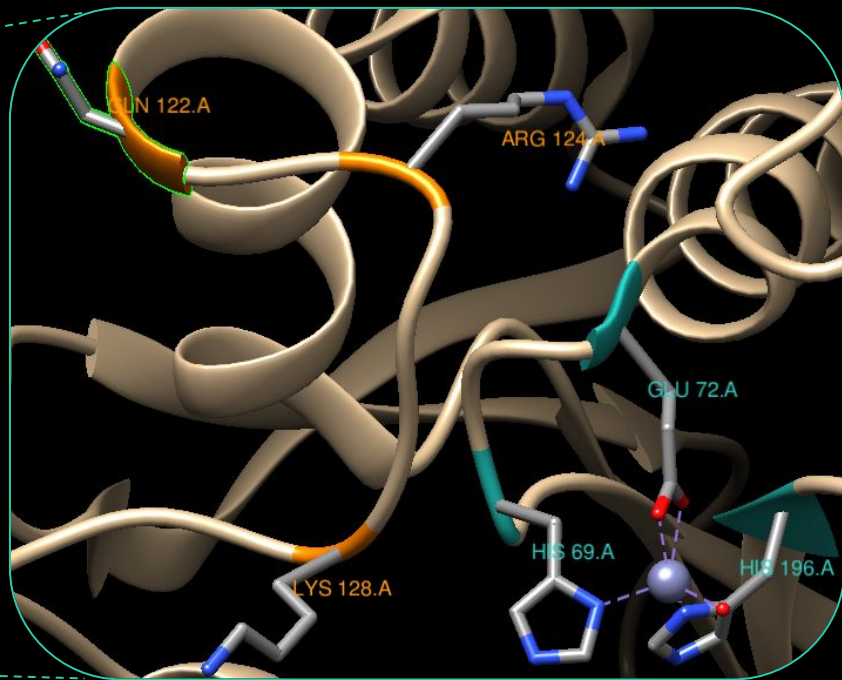
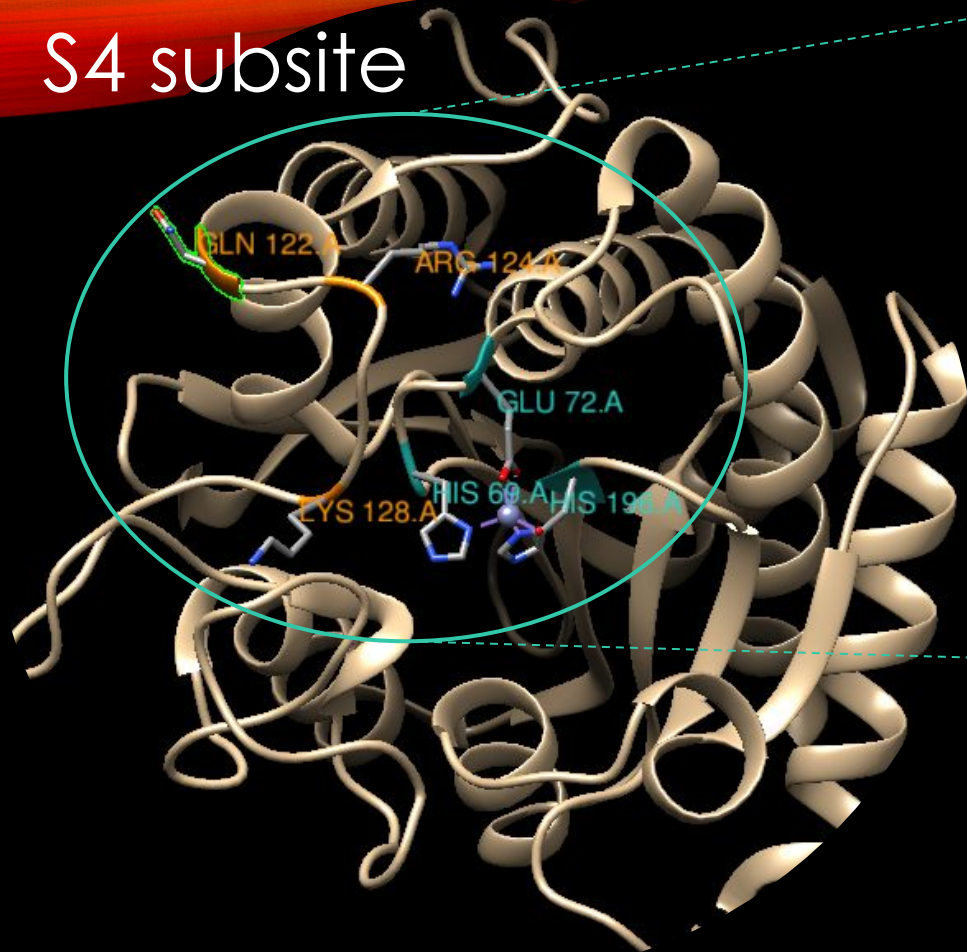


S3 subsite conservation



CBPA2_HUMAN	QASGGSIDWSYDYGIKYSFAFELRDTGRYGFLLPARQILPTAEETWLGLKAIMEHVDRDHP
CBPA1_RAT	QASGSTIDWTYSQGIKYSFTFELRDTGLRGFLLPASQIIPTAEETWLALLTIMDHTVKHP
CBPA1_MOUSE	QASGSTIDWTYSQGIKYSFTFELRDTGLRGFLLPASQIIPTAEETWLALLTIMDHTVKHP
CBPA1_BOVIN	QASGGSIDWSYNQGIKYSFTFELRDTGRYGFLLPASQIIPTAQETWLGVLTIMEHTLNHL
CBPA1_HUMAN	QASGSTIDWTYSQGIKYSFTFELRDTGRYGFLLPASQIIPTAKETWLALLTIMEHTLNHP
CBPA1_PIG	QASGGTIDWTYNQGIKYSFSFELRDTGRYGFLLPASQIIPTAQETWLALLTIMEHTLNHP
	****:***:* *****:***** ****** **:***:*****::*:*:..:

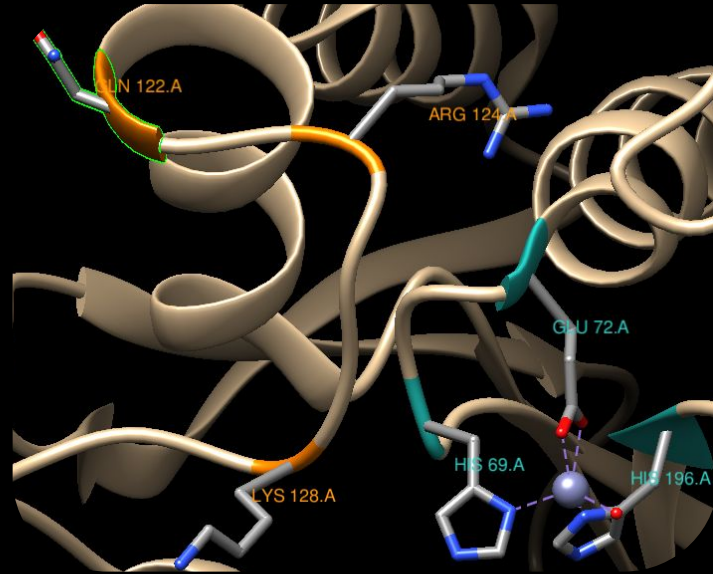
S4 subsite



S4 subsite alignment

Residue 122 is variable between species and between carboxypeptidases.

Despite this, the amino acid in this position conserves the polarity feature.



Gln 122 Arg 124 Lys 128

CBPA2_HUMAN	HAREWVTQATALWTANKIVSDYGKDPSITSILDALDIFLLPVTNPDGYVFSQTK	NRMWRK
CBPA1_RAT	HSREWVTQASGVWFAKKITKDYGQDPTFTAVLDNMDIFLEIVTNPDGFAYTHKT	NRMWRK
CBPA1_MOUSE	HSREWVTQASGVWFAKKITKDYGQEPTLTAILDNMDIFLEIVTNPDGFFVYTHKT	NRMWRK
CBPA1_BOVIN	HSREWITQATGVWFAKKFTEDYGQDPSFTAILDSMDIFLEIVTNPDGFAFTHSQ	NRLWRK
CBPA1_HUMAN	HSREWVTQASGVWFAKKITQDYGQDAAFTAILDTLDIFLEIVTNPDGFAFTHST	NRMWRK
CBPA1_PIG	HSREWVTQASGVWFAKKITEDYGQDPAFTAILDNLDIFLEIVTNPDGFAFTHSE	NRMWRK
	*:***:***:..* *:*:..***:::~::~** :***** *****:~::~	*~::~

Catalytic reaction

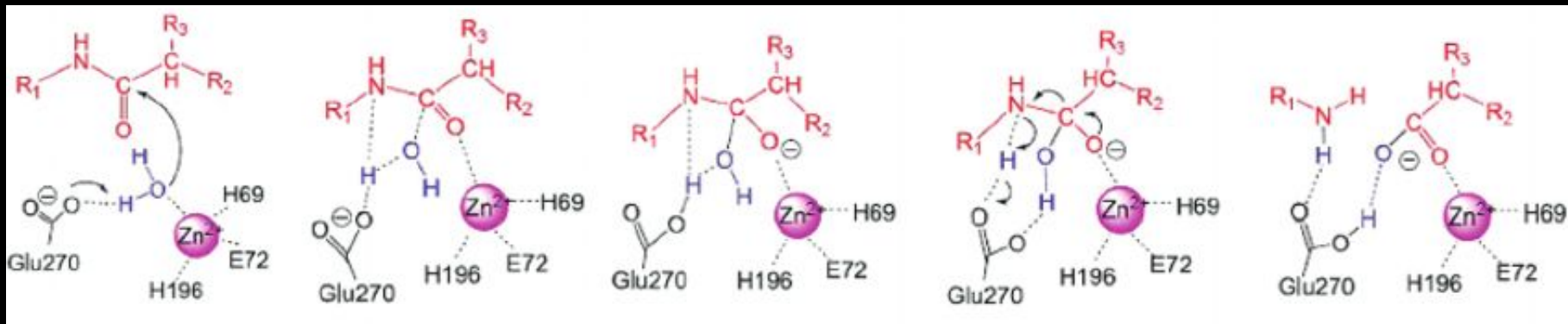
**2 POSSIBLE REACTIONS
STILL UNDER DEBATE**

PROMOTED WATER

**NUCLEOPHILIC OR ANHYDRIDE
PATHWAY**

Preferred mechanism

Promoted Water (general base-general acid)

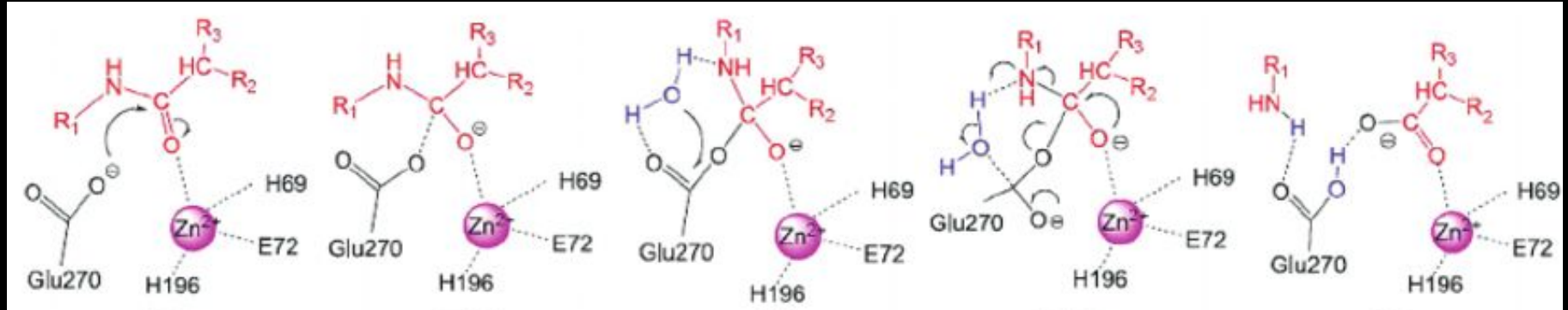


Wu S, Zhang C, Xu D, Guo H. Catalysis of Carboxypeptidase A: Promoted-Water versus Nucleophilic Pathways. *The Journal of Physical Chemistry B*. 2010;114(28):9259-9267.

STEP 1 Nucleophilic addition: Glu 270 acts as a general base facilitating the attack of the zinc-bound water at the scissile carbonyl carbon by transferring a water H⁺ to a carboxylate oxygen

STEP 2 Elimination: Glu 270 acts as a general acid where gives the H⁺ to the leaving nitrogen group

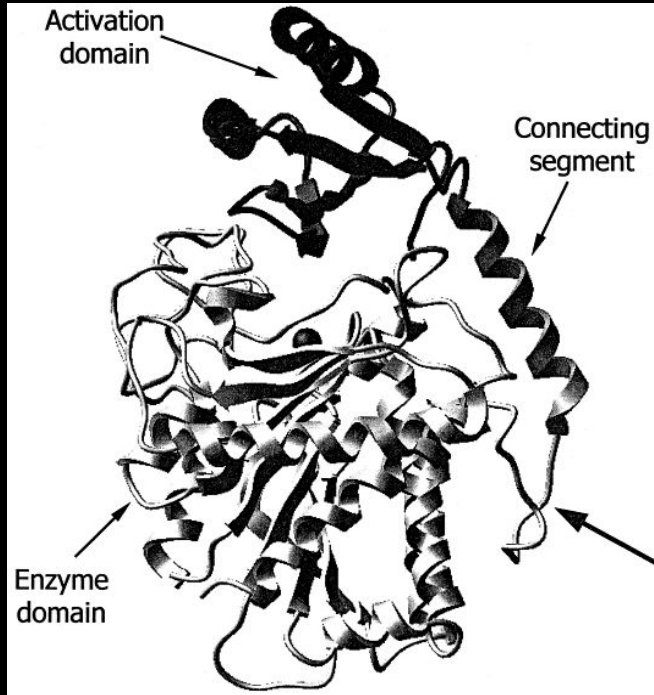
Anhydride or nucleophilic mechanism



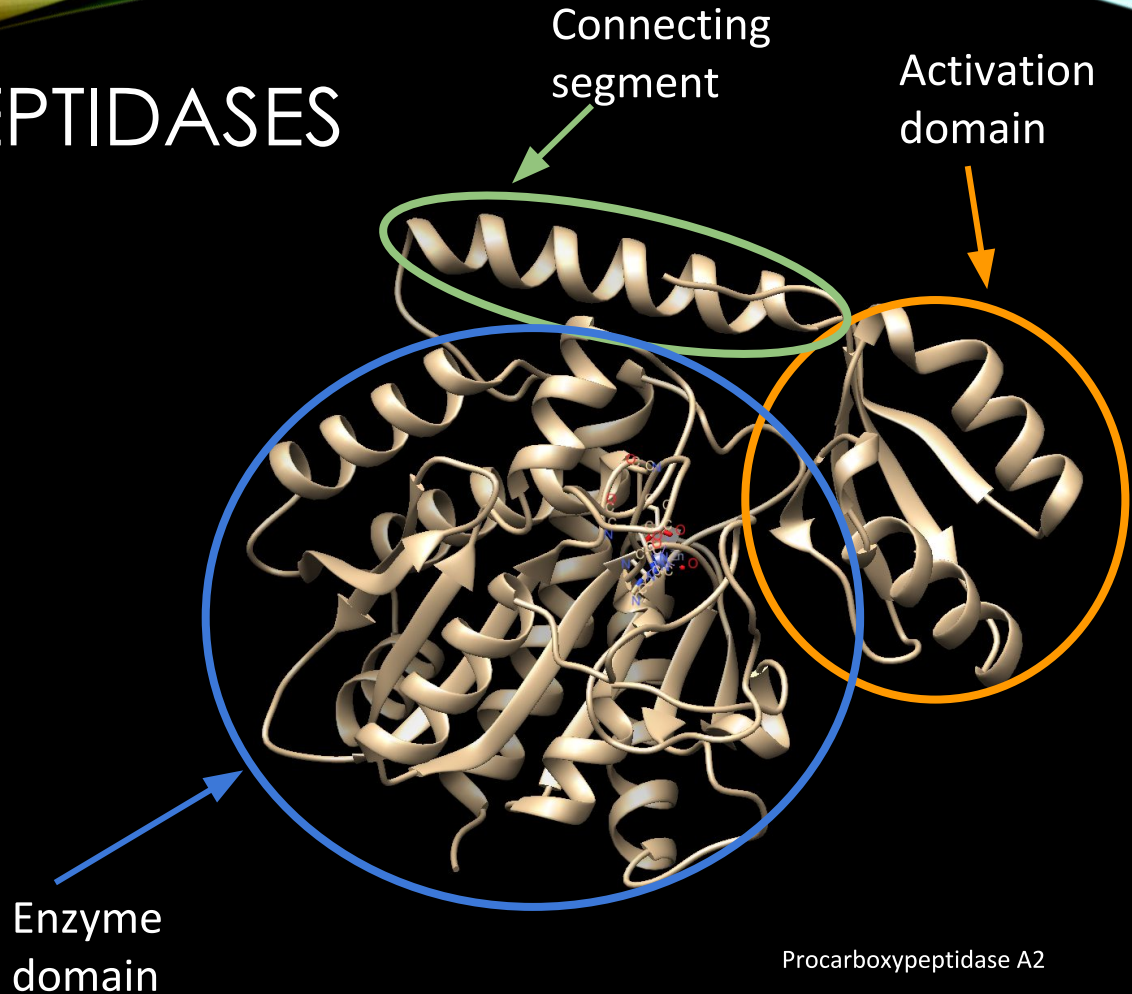
Wu S, Zhang C, Xu D, Guo H. Catalysis of Carboxypeptidase A: Promoted-Water versus Nucleophilic Pathways. *The Journal of Physical Chemistry B*. 2010;114(28):9259-9267.

Glu 270 performs a nucleophilic attack at the scissile carbonyl carbon, this needs an acyl-enzyme intermediate which can be posteriously hydrolysed by water.

PROCARBOXYPEPTIDASES

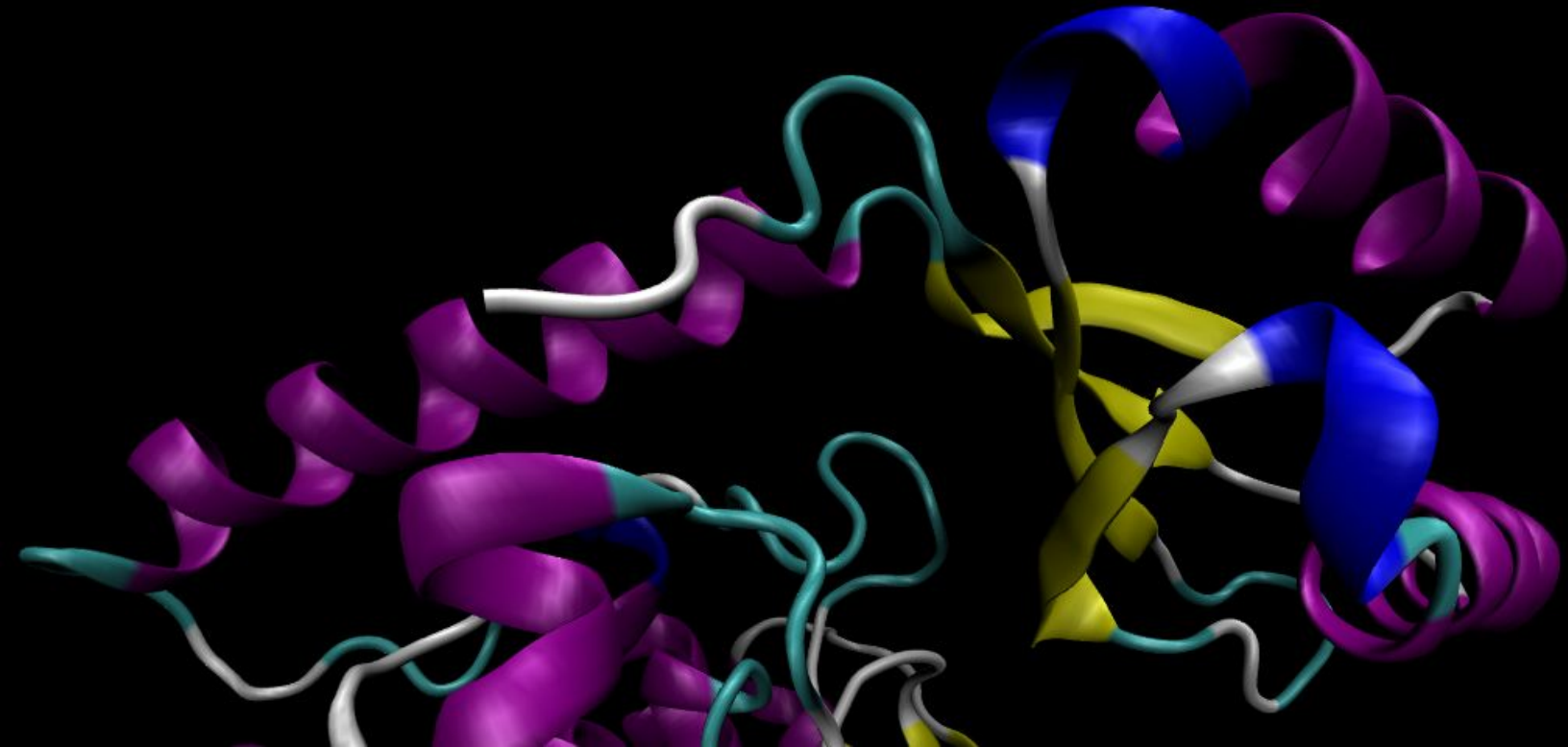


Vendrell J, Querol E, Avilés F. Metalloprocarboxypeptidases and their protein inhibitors. *Biochimica et Biophysica Acta (BBA) - Protein Structure and Molecular Enzymology*. 2000;1477(1-2):284-298.

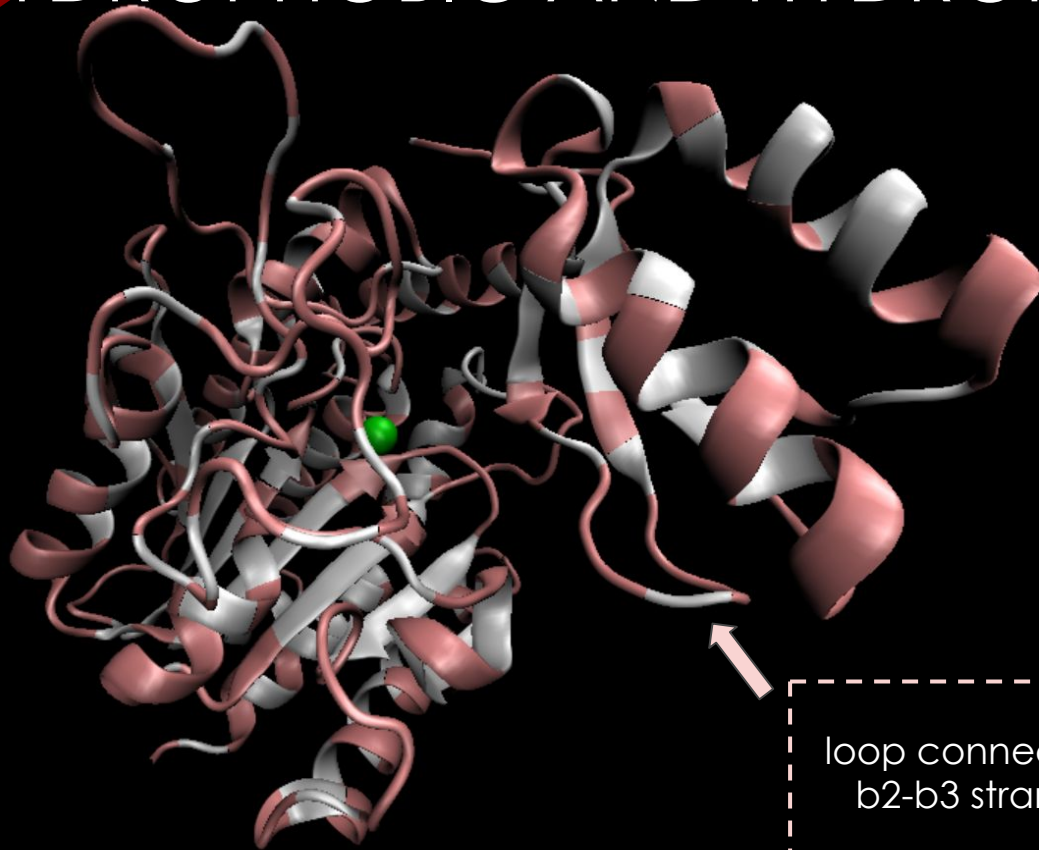


ACTIVATION DOMAIN

4 antiparallel β -strands
3 α helix
2 short 3_{10} helix
No disulfide bridges



HYDROPHOBIC AND HYDROPHILIC REGIONS



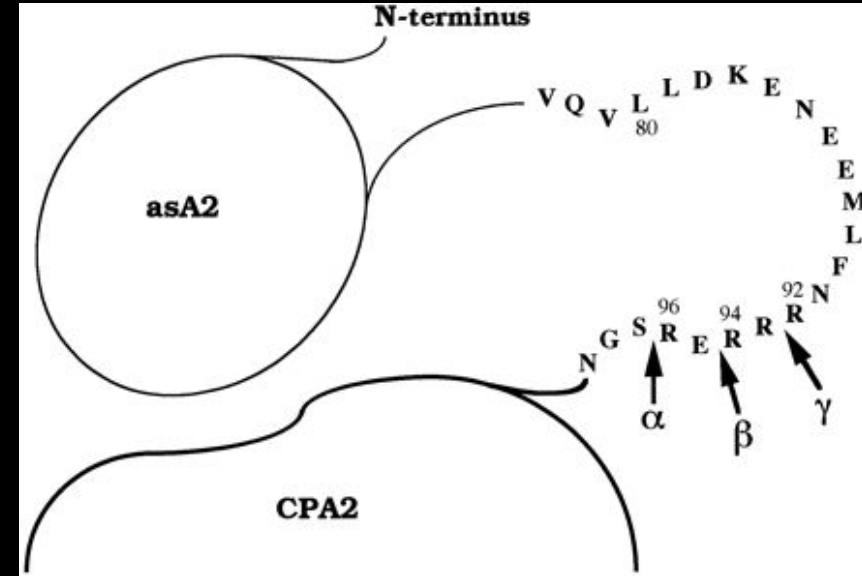
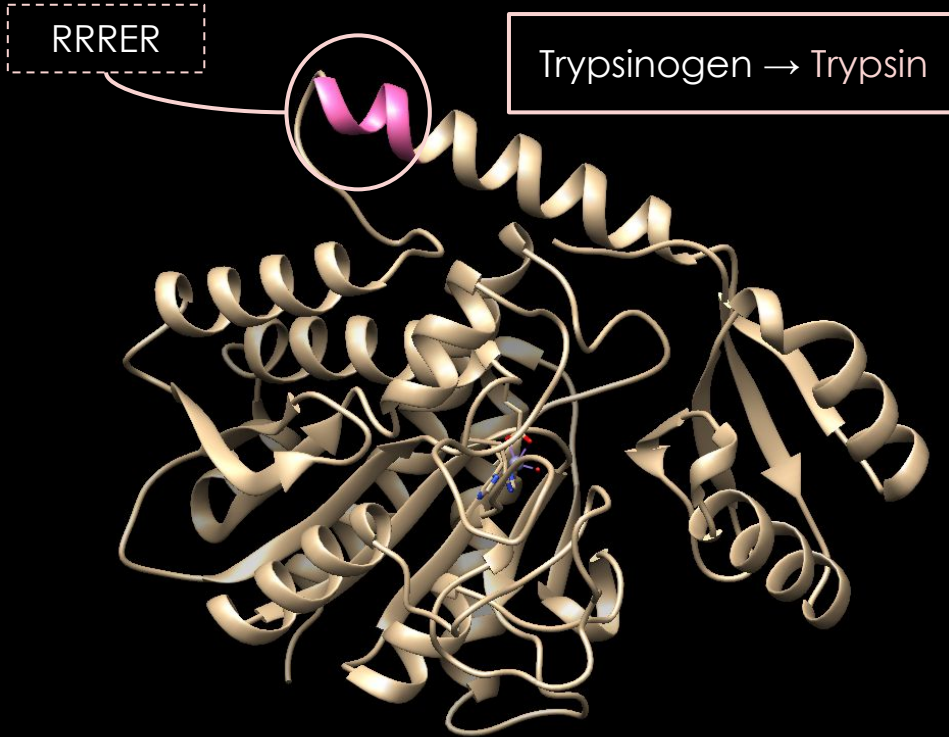
Hydrophobic residues

Hydrophilic residues

Zinc ion

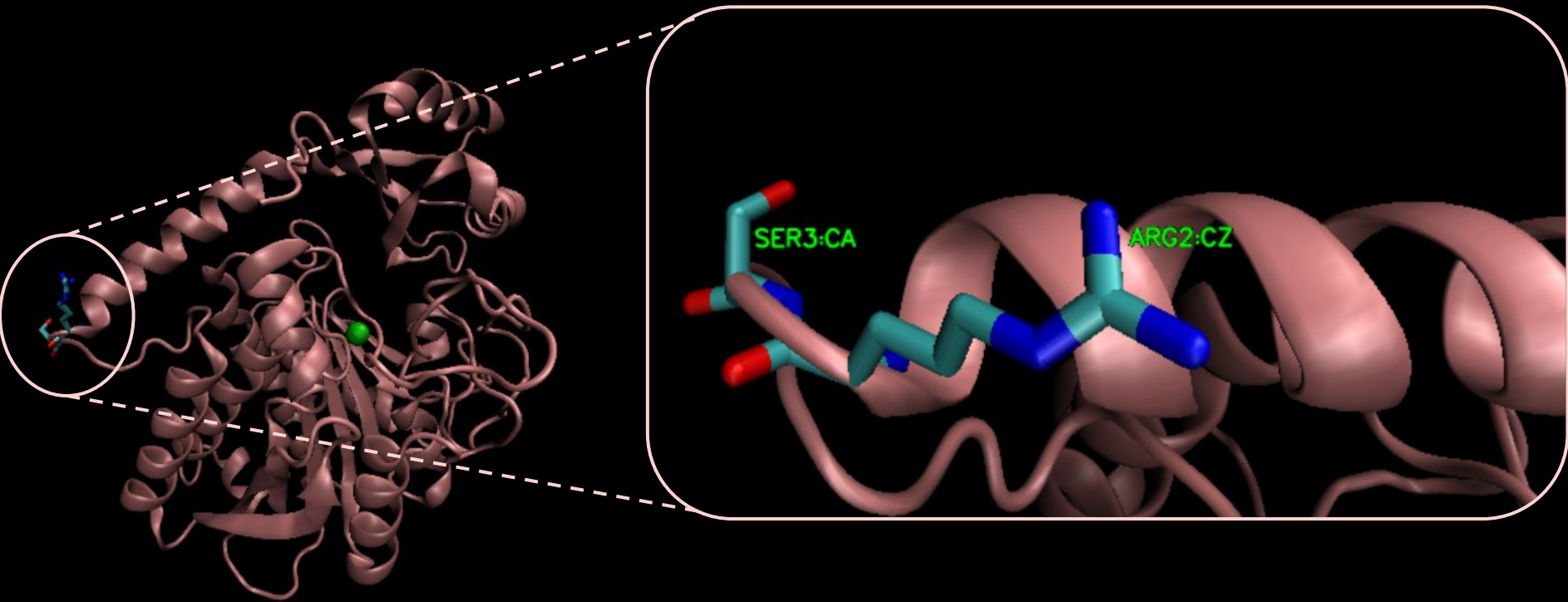
loop connecting
b2-b3 strands

TRYPSIN TARGET



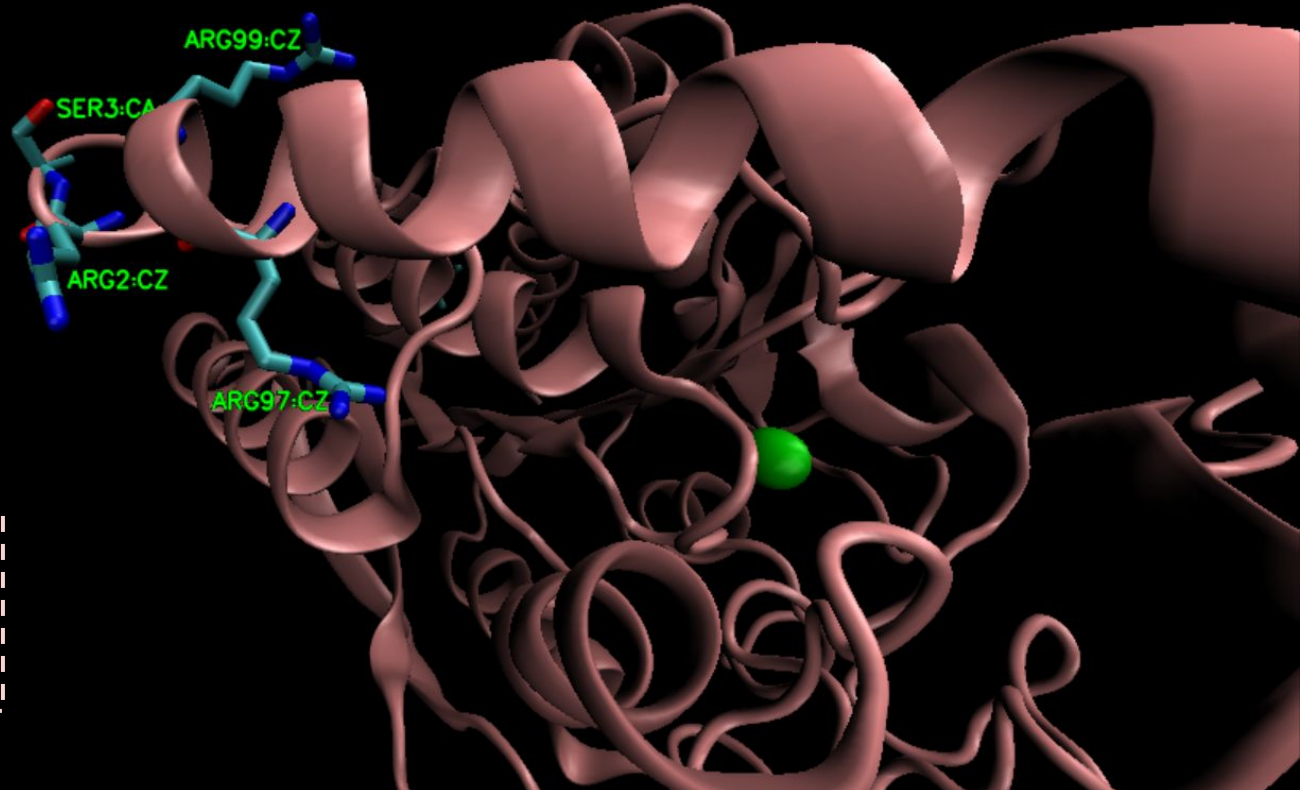
Reverter D, Ventura S, Villegas V, Vendrell J, Avilés F. Overexpression of Human Procarboxypeptidase A2 in *Pichia pastoris* and Detailed Characterization of Its Activation Pathway. *Journal of Biological Chemistry*. 1998;273(6):3535-3541.

TRYPSIN CLEAVAGE 1



TRYPSIN CLEAVAGE

1. **Arg2-Ser3**
2. Arg97-Arg99



- Porcine PCPA1 → 4 helix with Arg99
- Porcine PCPB → 2 helix

ALIGNMENT PCPA2-CPA2

PCPA2 → 1AYE
CPA2 → 1DTD

CLUSTAL O(1.2.4) multiple sequence alignment

```
1AYE      LETFVG DQVLEIVP SNEEQIKNLLQLEAQEHLQLDFWKSPTTPGETAHVRVPFVNVQAVK 60
1DTD      -----
1AYE      VFLESQGIAYSIMIEDVQVLLDKENEEMLFNRRRERSGNFNFNGAYHTLEEISQEMDNLVA 120
1DTD      ----- FNFNGAYHTLEEISQEMDNLVA 21
                  *****
1AYE      EHPGLVSKVNI GSSFENRPMNVLFKSTGGDKPAIWLDAGIHAREWVTQATALWTANKIVS 180
1DTD      EHPGLVSKVNI GSSFENRPMNVLFKSTGGDKPAIWLDAGIHAREWVTQATALWTANKIVS 81
                  *****
1AYE      DYGKDP SITSILDALDIFLLPVTNPDGYVFSQTKNRMWRKTRSKVS-GSLCVGVDPNRRNW 239
1DTD      DYGKDP SITSILDALDIFLLPVTNPDGYVFSQTKNRMWRKTRSKVSAGSLCVGVDPNRRNW 141
                  *****
1AYE      DAGFGGPGASSNPCSDSYHGPSANSEVEVKSI VDFIKSHGKVKAFIILHSYSQLLMFPYG 299
1DTD      DAGFGGPGASSNPCSDSYHGPSANSEVEVKSI VDFIKSHGKVKAFIILHSYSQLLMFPYG 201
                  *****
1AYE      YKCTKLDDFDELSEVAQKAAQSLRSLHGTKYKVGPICSVIYQASGGSIDWSYDYGIKYSF 359
1DTD      YKCTKLDDFDELSEVAQKAAQSLRSLHGTKYKVGPICSVIYQASGGSIDWSYDYGIKYSF 261
                  *****
1AYE      AFELRDTGRYGFLLPARQILPTAEETWLGLKAIMEHVRDHPY 401
1DTD      AFELRDTGRYGFLLPARQILPTAEETWLGLKAIMEHVRDHPY 303
                  *****
```


INTERACTIONS

García-Saez I, Reverter D, Vendrell J, Avilés F, Coll M. The three-dimensional structure of human procarboxypeptidase A2. Deciphering the basis of the inhibition, activation and intrinsic activity of the zymogen. The EMBO Journal. 1997;16(23):6906-6913.

Table 1. Interactions^a between the pro-segment and the CPA2 moiety in human pro-CPA2

Globular domain/CPA2	Distance (Å)	Connecting segment/CPA2	Distance (Å)
N(Glu5A)--O(Lys122)	3.0	C ⁷¹ (Val82A)...C ^{δ1} (Leu280)	4.0
O(Glu5A)--W2--N(Lys124)	3.0/2.9	C ^β (Leu85A)...C ^{δ1} (Leu280)	3.9
O(Glu5A)--W2--O(Lys124)	3.0/3.5	O(Leu85A)--W64--O(Leu280)	3.4/2.9
C ^{ε1} (Phe7A)...C ^γ (Met125)	3.7	C ^{δ1} (Leu86A)...C ^γ (Arg124)	3.8
C ^ε (Phe7A)...C ^γ (Met125)	3.7	O ^{ε1} (Glu89A)--N ^{η2} (Arg124)	2.9
O ^{ε2} (Glu33A)--N ^{δ2} (Asn159)	3.3	O ^{ε2} (Glu89A)--N ^ε (Arg124)	2.8
O ^{δ1} (Asp36A)--N ^{η2} (Arg71)	2.8	C ^δ (Glu89A)--C ^{ε3} (Trp73)	3.7
O ^{δ1} (Asp36A)--N ^{η1} (Arg71)	3.4	O ^{ε1} (Glu89A)--W65--O(Leu281)	2.8/2.9
O ^{δ2} (Arg36A)--N ^{η2} (Arg71)	3.0	O ^{ε2} (Glu92A)--N ^{η2} (Arg284)	3.3
O ^{δ1} (Asp36A)--W55--O ^{δ2} (Asp163)	2.7/2.8	O ^{ε1} (Glu92A)--W132--N ^ε (Ala283)	2.9/3.0
O(Trp38A)--W120--N ^{η1} (Arg71)	3.1/3.1	O ^{ε1} (Glu92A)--W100--N(Arg284)	2.8/2.9
C ^γ (Trp38A)...C ^{ε2} (Phe279) ^b	3.5	O ^{ε1} (Glu92A)--W00--N ^ε (Arg284)	2.8/2.9
C ^{ε3} (Trp38A)...C ^α (Gly278)	4.0	O(Met93A)--W87--O(Tyr12)	2.9/2.9
N ^ε (Lys39A)--W256--O(Val246)	2.8/2.7	O(Asn96A)--N(Glu1) ^c	2.8
N(Ser40A)--W13--O(Tyr248) ^f	3.3/3.2	N ^{δ2} (Asn96A)--O ^{ε2} (Glu1)	2.9
O(Ser40A)--W13--O(Tyr248) ^f	3.2/3.2	O ^{δ1} (Asn96A)--N ^{δ2} (Asn8)	3.0
C ^{γ2} (Thr42A)...C(Ile247)	3.9	C ^γ (Asn96A)--C ^β (Ala11)	3.6
C ^{γ2} (Thr42A)...C ^{ε2} (Tyr248) ^d	3.5	O(Arg97A)--N(Arg2) ^e	3.5
C ^{γ2} (Thr42A)...C ^β (Tyr248) ^c	3.8	O(Arg97A)--N(Glu1) ^e	3.3
C ^{γ2} (Thr47A)...C ^{ε2} (Tyr248) ^d	4.0	N ^{η2} (Arg97A)--W36--O(Tyr12)	2.7/3.5
N ^{δ1} (His53A)...O ^η (Tyr198)	2.7	Ne(Arg97A)--W36--O(Tyr12)	2.9
S ^δ (Met78A)...O(Gly275)	3.7	O(Arg98A)--N(Glu1) ^e	3.3
S ^δ (Met78A)...C(Gly275)	4.0	O(Arg98A)--N(Arg2) ^e	3.9
		O(Arg99A)--N(Gly4) ^e	3.1
		O(Arg99A)--N(Gly3) ^e	3.1
		O(Met93A)--W87--O(Tyr12)	2.9/2.9
		N ^{δ2} (Asn96A)--W87--O(Tyr12)	2.7/2.9
		N ^ε (Arg97A)--W36--O(Tyr12)	2.9/3.5
		N ^{η2} (Arg97A)--W36--O(Tyr12)	2.7/3.5

^a(...): van der Waals contacts ≤ 4.0 Å; (---): H-bonds ≤ 3.5 Å.

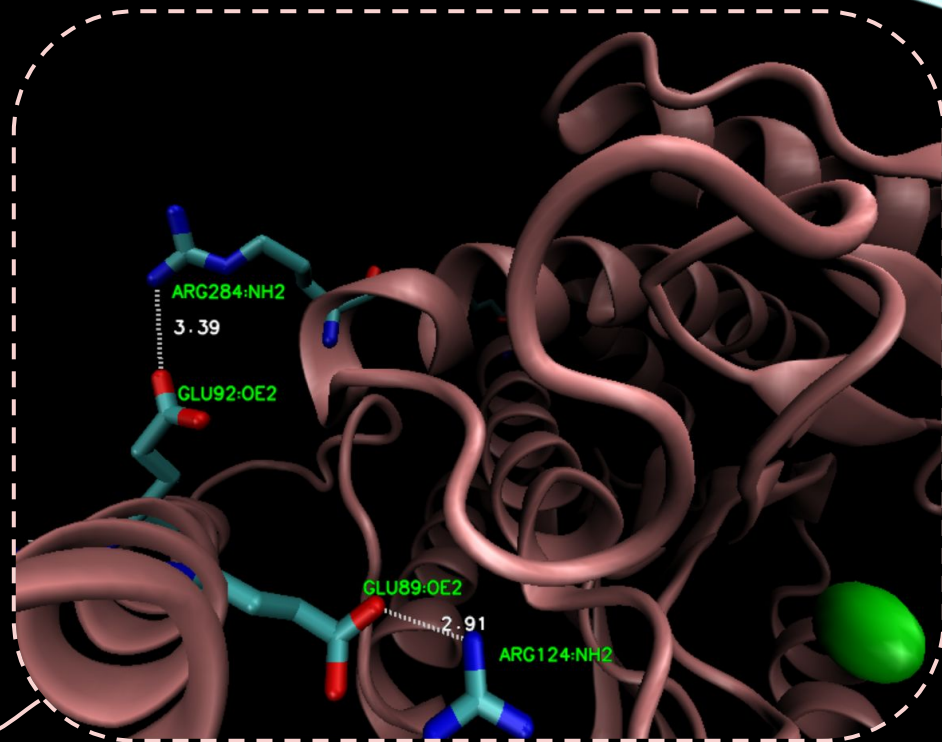
^bAromatic-aromatic interaction; only the shortest distance between the two rings is listed.

^cIn the benzylsuccinate complexed structure.

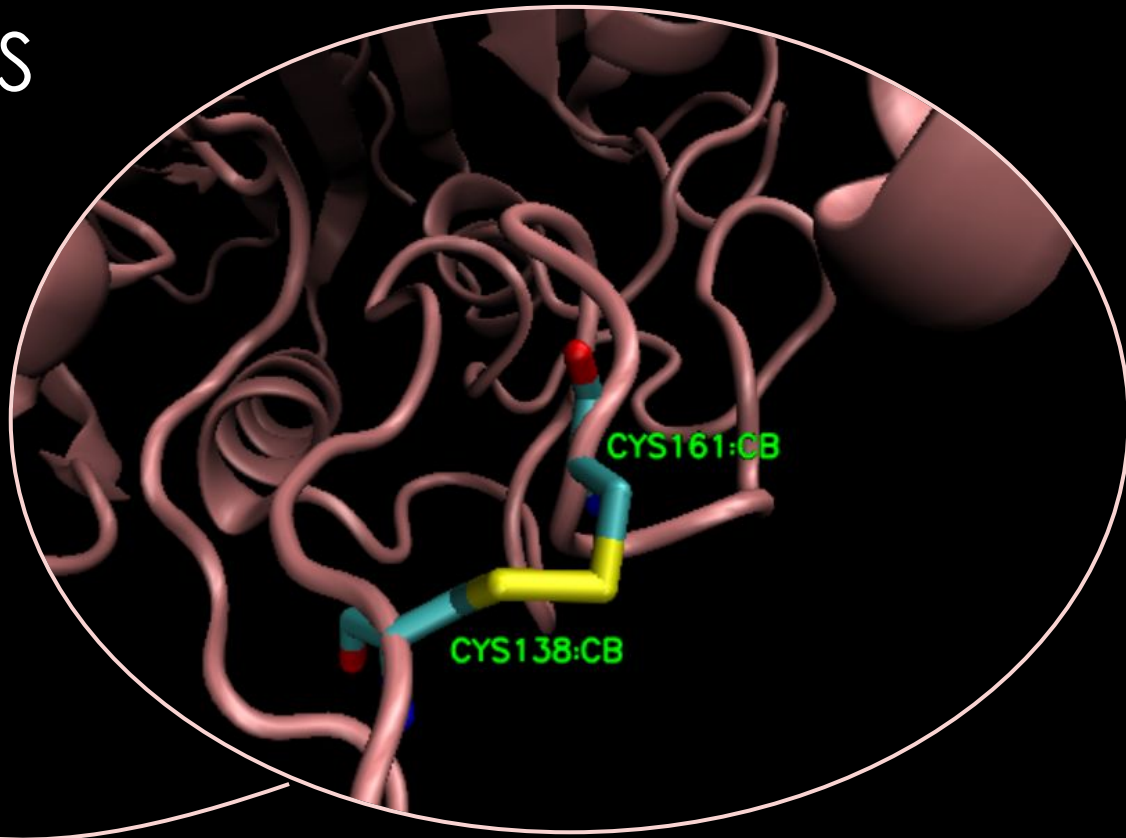
^dIn the uncomplexed structure.

^eIntrahelical H-bond.

SALT BRIDGES

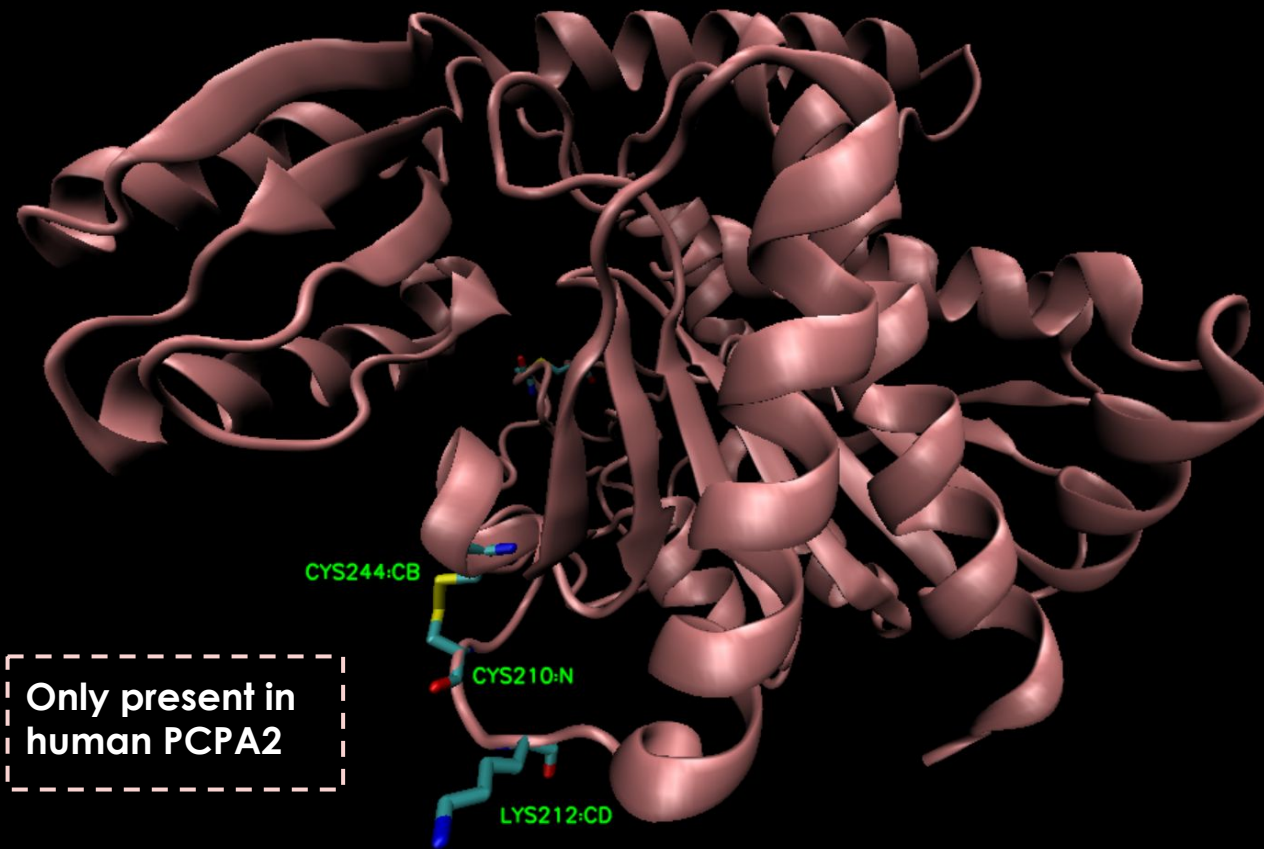


DISULFIDE BRIDGES



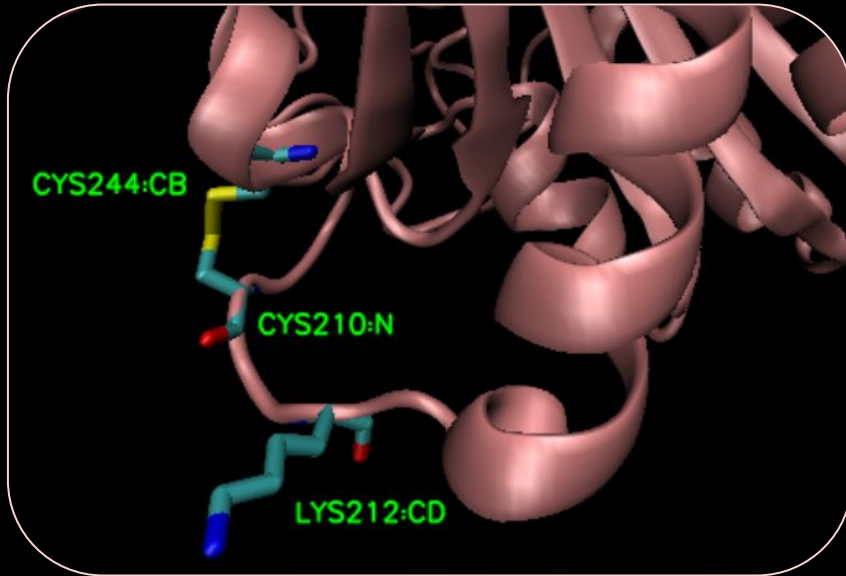
Present in porcine and bovine PCPA1

DISULFIDE BRIDGES

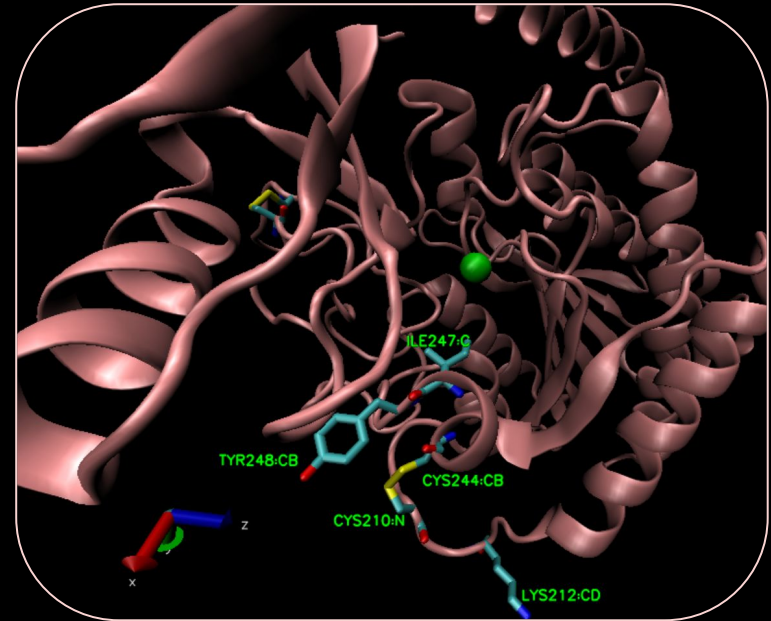


Only present in
human PCPA2

DISULFIDE BRIDGES

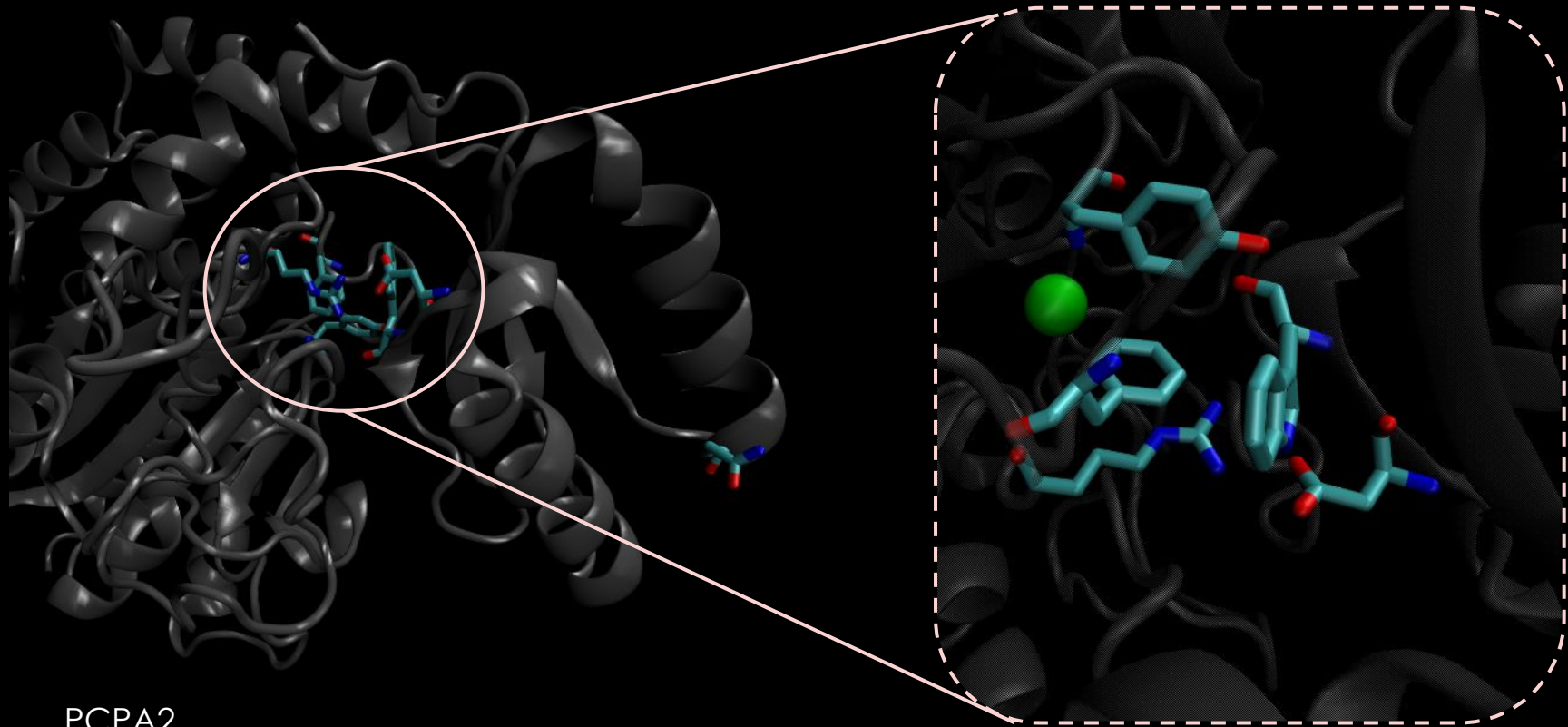


Displaces Lys
1.8Å



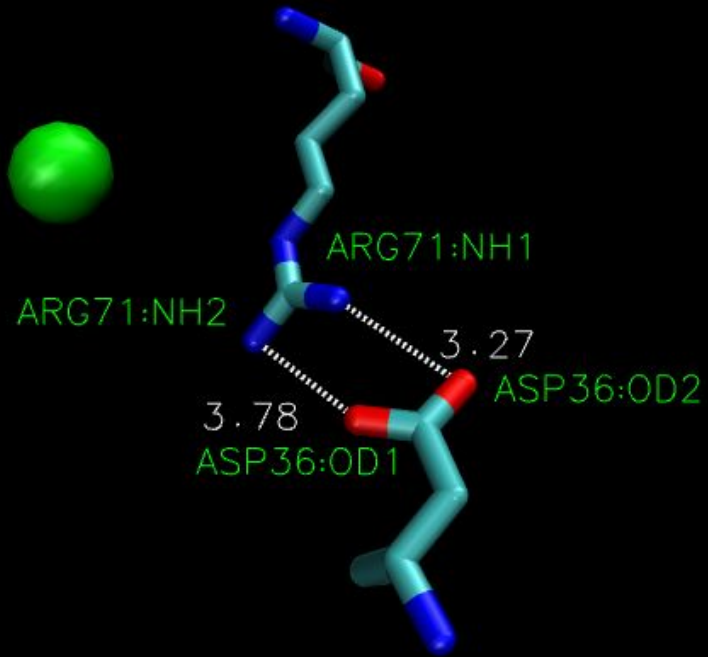
Displaces
residues 247-248

HYDROGEN BONDS

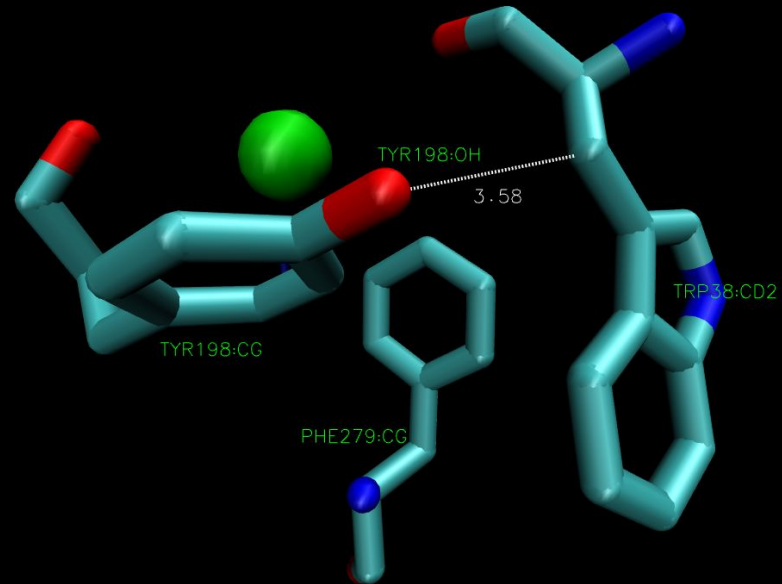


PCPA2

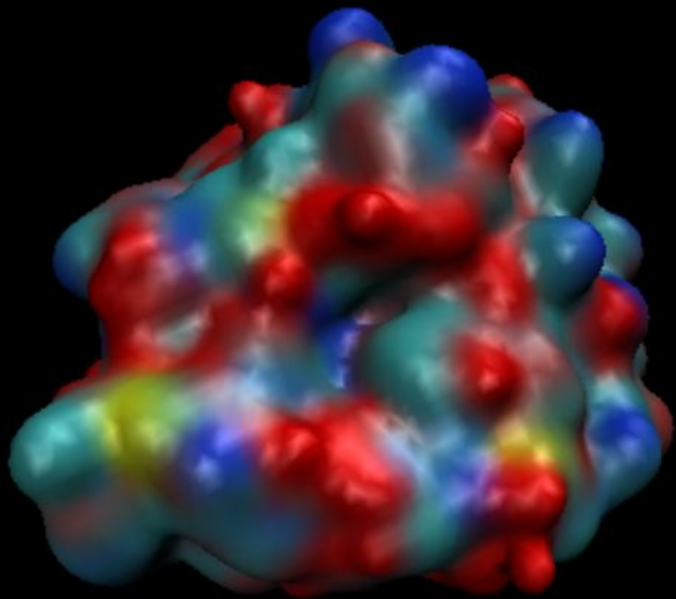
HYDROGEN BONDS



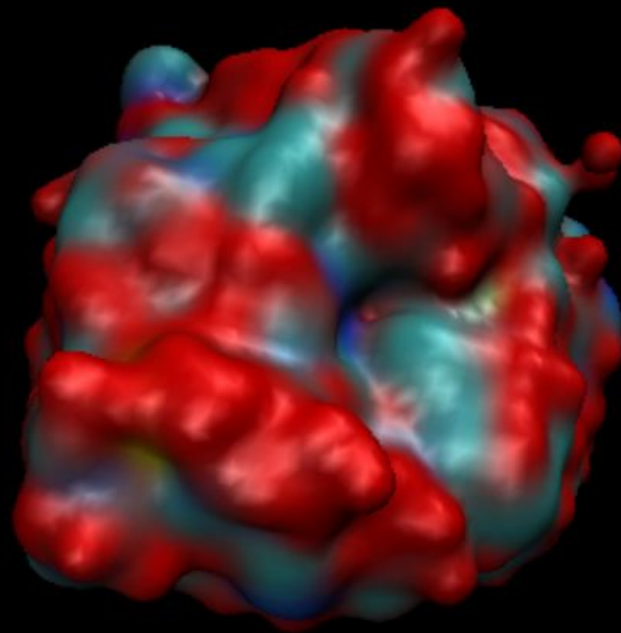
Pi-type interaction (Phe279-Trp38) -- 3.5



CARBOXYPEPTIDASE A VS B

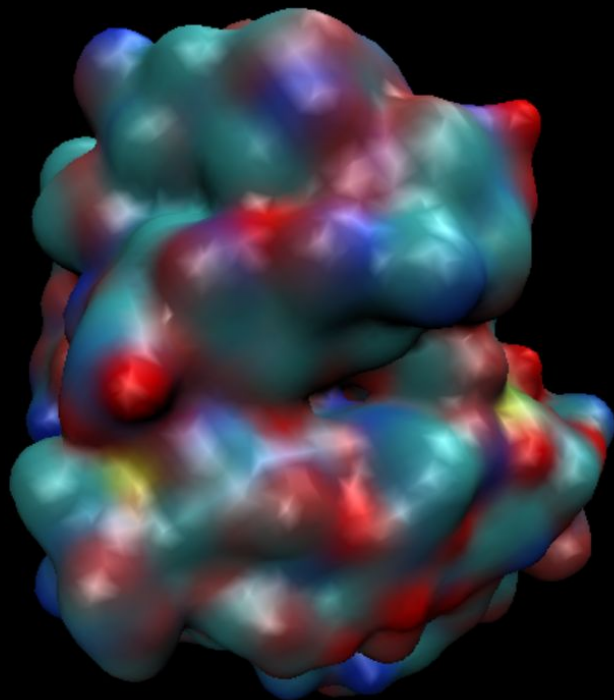


CPA2

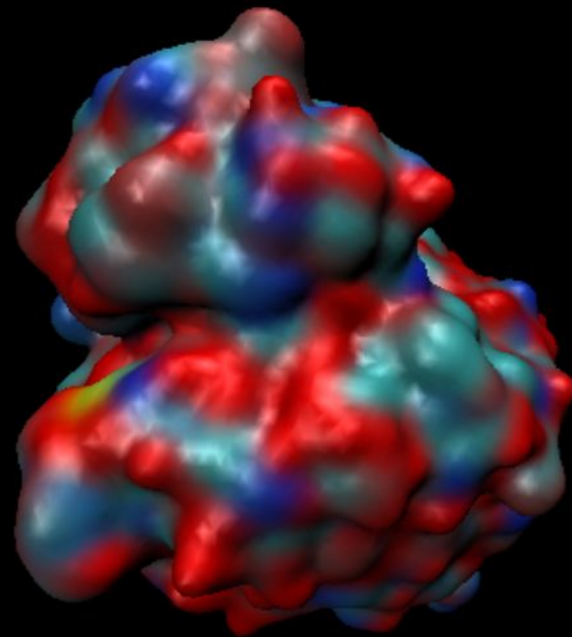


CPB

PROCARBOXYPEPTIDASE A VS B



PCPA2
Little activity



PCPB
No activity

TYPES OF CARBOXYPEPTIDASE INHIBITION



Autologous Inhibition

by Pro-Segment



Heterologous Inhibition

by Potato
Carboxypeptidase
Inhibitor

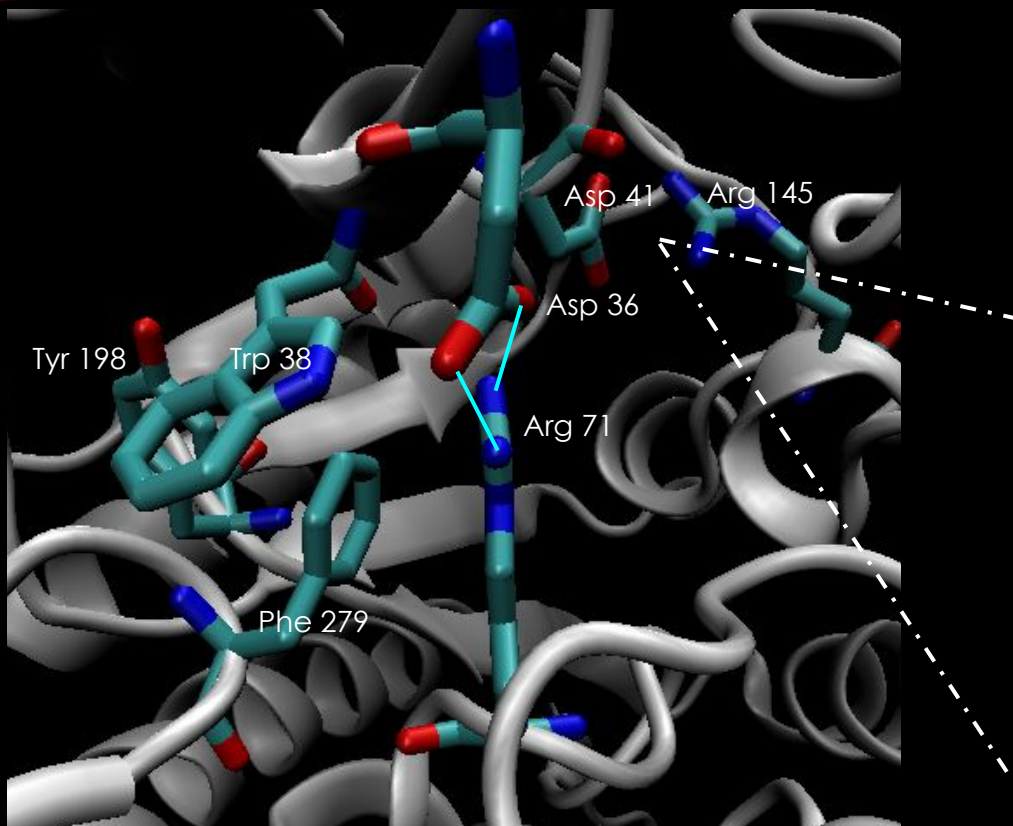
= PCI



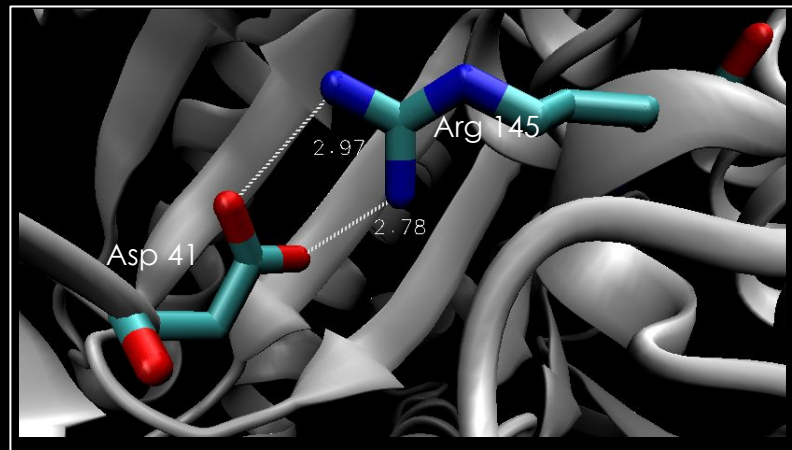
Excess of Zinc

Inhibition by a second
eq. of Zn^{2+}

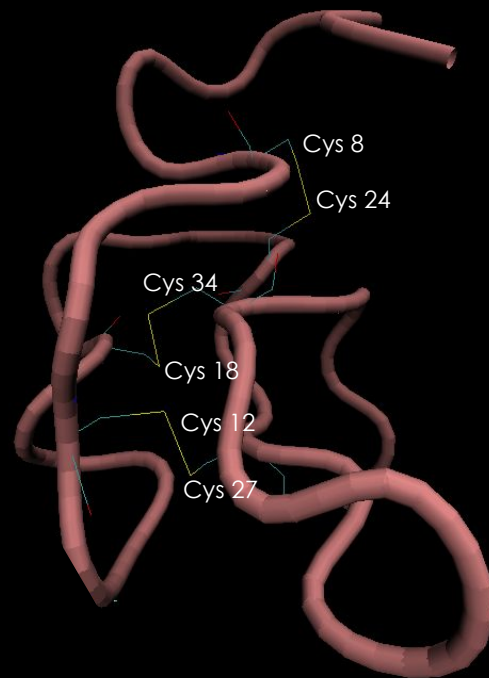
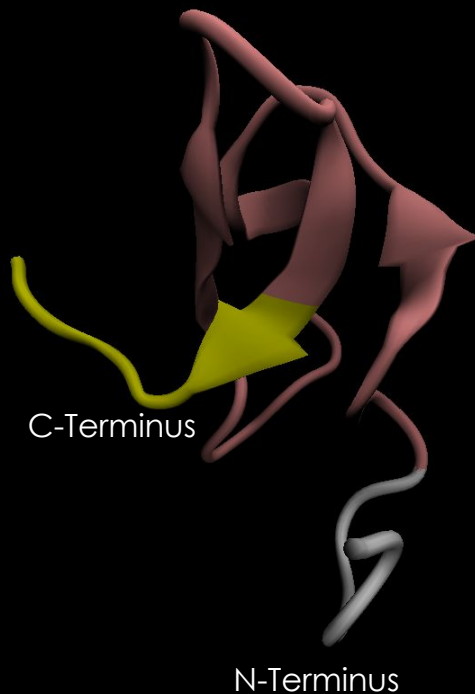
AUTOLOGOUS INHIBITORS: PRO-SEGMENT



salt bridge: Asp 41 - Arg 145

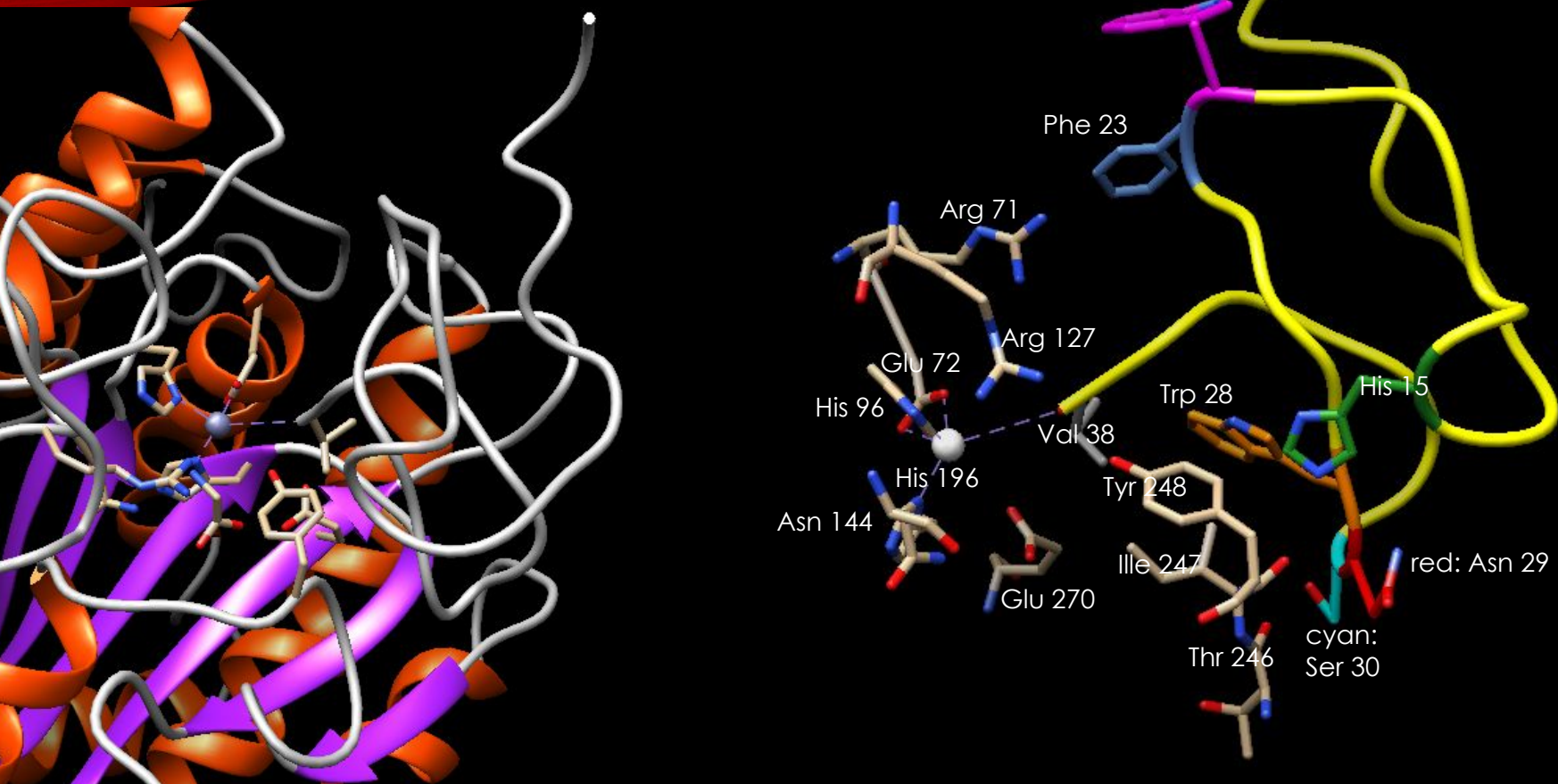


HETEROLOGOUS INHIBITORS: PCI

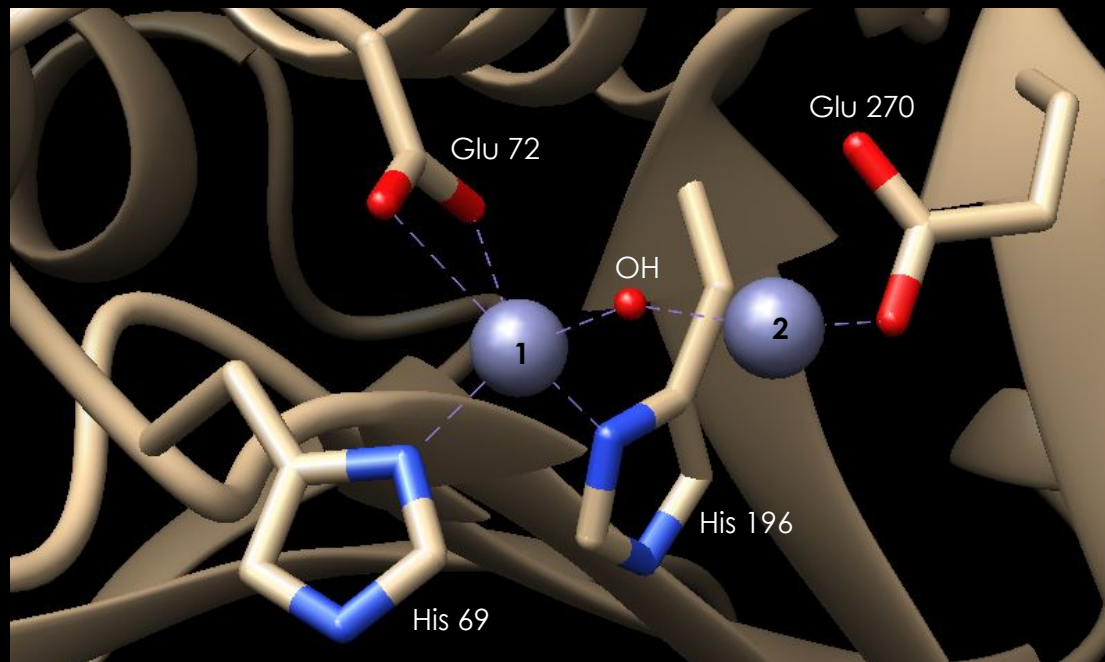


HETEROLOGOUS INHIBITORS: PCI

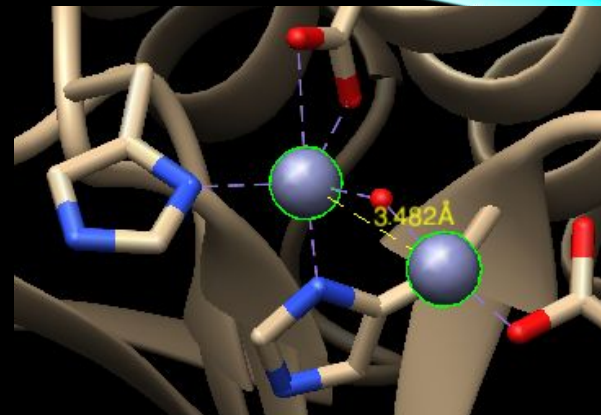
Interaction CPA-PCI:



EXCESS OF ZINC



- Larsen and Auld: Inhibition via Zincmonohydroxide (ZnOH^+)
- Zn^{2+} 1: catalytic Zn^{2+} 2: excess



atoms	distance in Å
Zn 1 - Zn 2	3.482
Zn 1 - OH	1.752
OH - Zn 2	1.846

torsion angle:

Zn 1 - OH - Zn 2: 150.887°

FURTHER INHIBITORS OF CARBOXYPEPTIDASES



Tomato Carboxypeptidase Inhibitor



Ascaris suum Carboxypeptidase Inhibitor
(parasit)



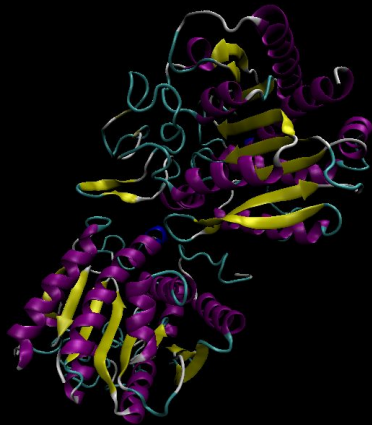
Medical leech Carboxypeptidase Inhibitor
(*Hirudo medicinalis*)



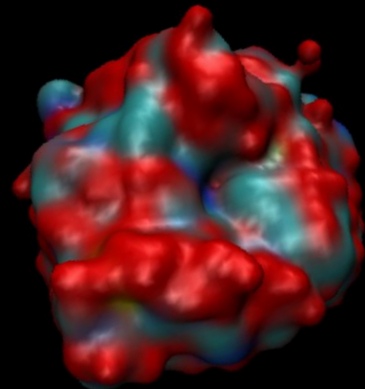
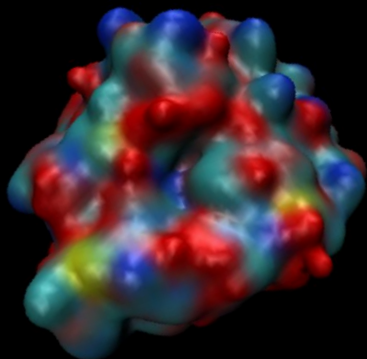
Rat brain Carboxypeptidase Inhibitor or tissue Carboxypeptidase
Inhibitor (TCI)

CONCLUSIONS

1. There are several kinds of CP's classifications
2. Carboxypeptidases belong to the exopeptidases
3. CPs act at C-terminals of the polypeptide chain of proteins and liberate a single amino acid or a dipeptide there
4. All of them show structural similarity in the catalytic domain
5. Pro-carboxypeptidases are the inactive form of carboxypeptidases and get activated by a trypsin cleavage of the pro-segment (96 aa).
6. There exist different ways of possible inhibitions
→ e.g. the autologous form occurs constantly in our organism
7. Play an important role in many processes such as digestion.



THANKS FOR YOUR ATTENTION!



REFERENCES

- Arolas J, Lorenzo J, Rovira A, Vendrell J, Aviles F, Ventura S. Secondary Binding Site of the Potato Carboxypeptidase Inhibitor. Contribution to Its Structure, Folding, and Biological Properties†. *Biochemistry*. 2004;43(24):7973-7982.
- Bech L, Soerensen S, Breddam K. Significance of hydrophobic S4-P4 interactions in subtilisin 309 from *Bacillus lentus*. *Biochemistry*. 1993;32(11):2845-2852.
- Banci L, Bertini I, La Penna G. The enzymatic mechanism of carboxypeptidase: A molecular dynamics study. *Proteins: Structure, Function, and Genetics*. 1994;18(2):186-197.
- Brändén C, Tooze J. Introduction to protein structure. New York, NY: Garland Pub.; 2009.
- Chapus C, Kerfelec B, Foglizzo E, Bonicel J. Further studies on the activation of bovine pancreatic procarboxypeptidase A by trypsin. *European Journal of Biochemistry*. 1987;166(2):379-385.
- Coll M, Guasch A, Avilés F, Huber R. Three-dimensional structure of porcine procarboxypeptidase B: a structural basis of its inactivity. *The EMBO Journal*. 1991;10(1):1-9.
- Deiteren K, Surpateanu G, Gilany K, Willemse J, Hendriks D, Augustyns K et al. The role of the S1 binding site of carboxypeptidase M in substrate specificity and turn-over. *Biochimica et Biophysica Acta (BBA) - Proteins and Proteomics*. 2007;1774(2):267-277.
- Fernández D, Avilés F, Vendrell J. Aromatic Organic Compounds as Scaffolds for Metallo-carboxypeptidase Inhibitor Design. *Chemical Biology & Drug Design*. 2009;73(1):75-82.
- Fernández Fleischhauer D, Avilés F. Discovery and characterization of small molecular weight metallo-carboxypeptidase inhibitors [Internet]. *Ddd.uab.cat*. 2019 [cited 26 February 2019]. Available from: <https://ddd.uab.cat/record/64748>

REFERENCES

Garcia-Guerrero M, Garcia-Pardo J, Berenguer E, Fernandez-Alvarez R, Barfi G, Lyons P et al. Crystal structure and mechanism of human carboxypeptidase O: Insights into its specific activity for acidic residues. *Proceedings of the National Academy of Sciences*. 2018;115(17):E3932-E3939.

Garcia-Saez I, Reverter D, Vendrell J, Avilés F, Coll M. The three-dimensional structure of human procarboxypeptidase A2. Deciphering the basis of the inhibition, activation and intrinsic activity of the zymogen. *The EMBO Journal*. 1997;16(23):6906-6913.

Gomez-Ortiz M, Gomis-Rüth F, Huber R, Avilés F. Inhibition of carboxypeptidase A by excess zinc: analysis of the structural determinants by X-ray crystallography. *FEBS Letters*. 1997;400(3):336-340.

Guasch A, Coll M, Avilés F, Huber R. Three-dimensional structure of porcine pancreatic procarboxypeptidase A. *Journal of Molecular Biology*. 1992;224(1):141-157.

Kim D, Shin Y, Kim K. The structural feature of S1' subsite of carboxypeptidase A. *Bioorganic & Medicinal Chemistry Letters*. 1991;1(6):317-322.

Laethem R, Blumenkopf T, Cory M, Elwell L, Moxham C, Ray P et al. Expression and Characterization of Human Pancreatic Preprocarboxypeptidase A1 and Preprocarboxypeptidase A2. *Archives of Biochemistry and Biophysics*. 1996;332(1):8-18.

Li X, Solomon B. Zinc-mediated thermal stabilization of carboxypeptidase A. *Biomolecular Engineering*. 2001;18(4):179-183.

MEROPS - the Peptidase Database [Internet]. Ebi.ac.uk. 2019 [cited 22 February 2019]. Available from: <https://www.ebi.ac.uk/merops/cgi-bin/famsum?family=M14>

Neurath H, Walsh K. Role of proteolytic enzymes in biological regulation (a review). *Proceedings of the National Academy of Sciences*. 1976;73(11):3825-3832.

REFERENCES

Pallarès I, Fernández D, Comellas-Bigler M, Fernández-Recio J, Ventura S, Avilés F et al. Direct interaction between a human digestive protease and the mucoadhesive poly(acrylic acid). *Acta Crystallographica Section D Biological Crystallography*. 2008;64(7):784-791.

Pfam: Family: Peptidase_M14 (PF00246) [Internet]. Pfam.xfam.org. 2019 [cited 22 February 2019]. Available from: https://pfam.xfam.org/family/Peptidase_M14

Prasad N. *Enzyme technology*. New Delhi: PHI Learning; 2011.

Reverter D, Ventura S, Villegas V, Vendrell J, Avilés F. Overexpression of Human Procarboxypeptidase A2 in *Pichia pastoris* and Detailed Characterization of Its Activation Pathway. *Journal of Biological Chemistry*. 1998;273(6):3535-3541.

Rees D, Lewis M, Honzatko R, Lipscomb W, Hardman K. Zinc environment and cis peptide bonds in carboxypeptidase A at 1.75-Å resolution. *Proceedings of the National Academy of Sciences*. 1981;78(6):3408-3412.

SCOP: Superfamily: Zn-dependent exopeptidases [Internet]. Scop.mrc-lmb.cam.ac.uk 2019 [cited 22 February 2019]. Available from: <http://scop.mrc-lmb.cam.ac.uk/scop/data/scop.b.d.hi.f.html>

Solomon B, Larsen K, Riordan J. Catalytic and conformational changes induced by limited subtilisin cleavage of bovine carboxypeptidase A. *Biochemistry*. 1990;29(31):7303-7309.

Tanco S, Lorenzo J, Garcia-Pardo J, Degroove S, Martens L, Aviles F et al. Proteome-derived Peptide Libraries to Study the Substrate Specificity Profiles of Carboxypeptidases. *Molecular & Cellular Proteomics*. 2013;12(8):2096-2110.

Vendrell J, Querol E, Avilés F. Metallo-carboxypeptidases and their protein inhibitors. *Biochimica et Biophysica Acta (BBA) - Protein Structure and Molecular Enzymology*. 2000;1477(1-2):284-298.

Wu S, Zhang C, Xu D, Guo H. Catalysis of Carboxypeptidase A: Promoted-Water versus Nucleophilic Pathways. *The Journal of Physical Chemistry B*. 2010;114(28):9259-9267.

PEM QUESTIONS

1. Which is correct about carboxypeptidases:
 - a. Procarboxypeptidase's prosegment is usually short, about 40aa.
 - b. Procarboxypeptidases have the same conformation as carboxypeptidases but with an activation domain or prosegment**
 - c. Carboxypeptidases and procarboxypeptidases have a different folding on the catalytic center.
 - d. Procarboxypeptidases are all beta proteins.
 - e. Carboxypeptidases have a selenium ion on its catalytic center

2. Which is the correct?
 - a. All procarboxypeptidases are fully inactive.
 - b. Procarboxypeptidases A1 and B have little activity.
 - c. Procarboxypeptidase A2 is fully inactive.
 - d. The trypsin target has lots of Asp.
 - e. All the answers are incorrect.**

PEM QUESTIONS

3. Which of the following examples can act as Inhibitors of CPs:

- a. Potato Carboxypeptidase inhibitor (PCI)
- b. Pro-segments
- c. Tomato carboxypeptidase inhibitor
- d. Only a and b are correct
- e. **All the answers are correct**

4. The pro-segments acts as inhibitors until ... :

- a. A second zinc-ion binds to the catalytic zinc-ion
- b. **Trypsin activation occurs**
- c. They can't act as inhibitors
- d. Tyrosine activation occurs
- e. None of this answers is correct

5. Why are His 96, His196 and Glu 72 so important residues?

- a. **They are zinc-coordinated**
- b. They don't have an important role
- c. Because they have positive charge
- d. Because they are aromatic residues
- e. They are residues in the sidechains which are responsible for the correct protein conformation

PEM QUESTIONS

6. Carboxypeptidases are ...?

- a. Endopeptidases which cleave one amino acid from the C-terminal end of the polypeptide substrates
- b. Endopeptidases which cleave one amino acid from the N-terminal end of the polypeptide substrates
- c. Exopeptidases which cleave one amino acid from the N-terminal end of the polypeptide substrates
- d. **Exopeptidases which cleave one amino acid from the C-terminal end of the polypeptide substrates**
- e. No answer of above is correct

7. Which metal ion uses the metalloenzyme CPA?

- a. Fe^{2+}
- b. Mn^{2+}
- c. **Zn^{2+}**
- d. Mg^{2+}
- e. Cu^{2+}

PEM QUESTIONS

8. Which sentence about procarboxypeptidases is correct?
- a. They are secreted in the liver
 - b. They need activation by trypsin**
 - c. The pro-peptide is about 60 residues long
 - d. They release the pro-peptide when they were cut by histidine
 - e. All answers are correct
9. Which of the following answers is correct? (CP=carboxypeptidase)
- a. Only CP A is secreted as a zymogen
 - b. Only CP B is secreted as a zymogen
 - c. Both are secreted as zymogens**
 - d. CP B doesn't exist
 - e. None of the answers is correct

PEM QUESTIONS

10. How many subsites has Carboxypeptidase A?

- a. 0
- b. 1
- c. 5**
- d. 3
- e. 2

MATERIALS AND METHODS

Programs:

Chimera
VMD
Clustalw
STAMP

Databases:

PDB
Uniprot
PFAM

